

AS17: Kapitel 7.6 - 7.8 Hypothesentest

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Contents

Aktuelle R-Umgebung zu den Beispielen	2
7.6 Mehrstichprobenverfahren	3
7.6.1 Prüfung der Gleichheit mehrerer Varianzen	3
7.6.3 Varianzanalyse (ANOVA)	4
7.6.4 Multiple Vergleiche	6
7.6.5 H-Test von Kruskal und Wallis	15
7.6.6 Varianzanalyse für Messwiederholungen	19
7.6.7 Friedman-Test	19
7.6.8 Zweifache Varianzanalyse	26
7.6.9 Analyse von wiederholten Messungen	28
7.6.10 Versuchsplanung	32
7.7 Die Analyse von Häufigkeiten	33
7.7.1 Vergleich relativer Häufigkeiten	33
7.7.2 Analyse von Vierfeldertafeln	35
7.7.3 Spezielle Risiko- und Effektmaße	38
7.7.4 Exakter Test nach R.A. Fisher	42
7.7.5 Äquivalenztest zweier Binomialwahrscheinlichkeiten	43
7.7.6 McNemar Vorzeichentest	44
7.7.7 Test nach Mantel-Haenszel	47
7.7.8 Der $k \times 2$ -Felder-Test nach Brandt und Snedecor	51
7.7.9 Cochran-Armitage-Test auf linearen Trend	54
7.7.10 Vergleich mehrerer Anteile mit einem Standard	56
7.7.11 Die Analyse von Kontingenztafeln	57
7.7.12 Bowker-Test auf Symmetrie	61
7.7.13 Marginalhomogenitätstest nach Lehman	61

7.7.14 Stuart-Maxwell-Test auf Homogenität in den Randverteilungen	63
7.7.15 Q-Test nach Cochran	64
7.7.16 Cohens's Kappa-Koeffizient	65
7.7.17 Krippendorff's Alpha	68
7.7.18 Kendall's Konkordanzkoeffizient	68
7.8 Hypothesentests zur Korrelation und Regression	69
7.8.1 Korrelationskoeffizient nach Pearson	69
7.8.2 Prüfung der Rangkorrelationskoeffizienten	75
7.8.4 Hypothesentests zu den Regressionskoeffizienten	77

Hinweis: Die Gliederung zu den Befehlen Und Ergebnissen in R orientiert sich an dem Inhaltsverzeichnis der 17. Auflage der 'Angewandten Statistik'. Nähere Hinweise zu den verwendeten Formeln sowie Erklärungen zu den Beispielen sind in dem Buch (Ebook) nachzulesen!

zur Druckversion

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Aktuelle R-Umgebung zu den Beispielen

The R Project for Statistical Computing

```
sessionInfo()
## R version 3.6.3 (2020-02-29)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 18363)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=German_Germany.1252 LC_CTYPE=German_Germany.1252
## [3] LC_MONETARY=German_Germany.1252 LC_NUMERIC=C
## [5] LC_TIME=German_Germany.1252
##
## attached base packages:
## [1] stats      graphics  grDevices  utils      datasets  methods   base
##
## loaded via a namespace (and not attached):
## [1] compiler_3.6.3  magrittr_1.5    tools_3.6.3    htmltools_0.4.0
## [5] yaml_2.2.1      Rcpp_1.0.4      stringi_1.4.6  rmarkdown_2.1
## [9] knitr_1.28      stringr_1.4.0   xfun_0.12      digest_0.6.25
## [13] rlang_0.4.5     evaluate_0.14
```

Download von Datensätzen, die in den folgenden Beispielen verwendet werden:

Wiederholte Messungen: repeated

Übereinstimmung (Bland-Altman): blandcor

7.6 Mehrstichprobenverfahren

7.6.1 Prüfung der Gleichheit mehrerer Varianzen

7.6.1.3 Bartlett-Verfahren

```
x <- c( 9, 11,  6, 11, 14,  7,  7, 11)           # Bartlett-Test
y <- c(13, 10, 12, 16, 11, 13, 15,  9,  9, 10)
z <- c( 7, 27,  8, 11, 17,  2, 16, 15,  9, 15, 18, 12)

k <- 3
si <- c(sd(x), sd(y), sd(z)); si
## [1] 2.725541 2.440401 6.444989
nui <- c(length(x)-1, length(y)-1, length(z)-1); nu <- sum(nui)
c <- (sum(1/nui) - 1/nu)/(3*(k-1)) + 1
ssqr <- sum(nui*si^2)/nu
chisqr <- 1/c*(2.3026 * (nu*log10(ssqr)-sum(nui*log10(si^2)))); chisqr
## [1] 10.36702
qchisq(0.95, k-1)
## [1] 5.991465
pchisq(chisqr, k-1, lower.tail=F)
## [1] 0.005608289

bartlett.test(list(x,y,z))
##
## Bartlett test of homogeneity of variances
##
## data:  list(x, y, z)
## Bartlett's K-squared = 10.367, df = 2, p-value = 0.005608
```

7.6.1.4 Levene-Test (Brown-Forsythe-Version)

Funktionen `leveneTest()` in `library(car)`

```
library(car)
val <- c(x, y, z)
grp <- as.factor(c(rep("I", length(x)),
                    rep("II", length(y)), rep("III", length(z))))

leveneTest(val ~ grp)                                # Levene-test
```

	Df	F value	Pr(>F)
group	2	3.904317	0.0324068
	27	NA	NA

```
fligner.test(val ~ grp)                                # Fligner-Test
##
## Fligner-Killeen test of homogeneity of variances
##
## data: val by grp
## Fligner-Killeen:med chi-squared = 7.3235, df = 2, p-value = 0.02569
```

7.6.3 Varianzanalyse (ANOVA)

```
gruppe <- c(1, 1, 2, 2, 2, 2, 3, 3, 3)                # Funktion aov()
wert <- c(3, 7, 4, 2, 7, 3, 8, 4, 6)
daten <- data.frame(gruppe=factor(gruppe), wert);

summary(aov(wert ~ gruppe, data=daten))
##              Df Sum Sq Mean Sq F value Pr(>F)
## gruppe        2  6.889   3.444   0.689  0.538
## Residuals     6 30.000   5.000
```

Beispiel (gleichgroße Stichprobenumfänge):

```
gruppe <- c(rep(1,4), rep(2,4), rep(3,4))              # Beispiel
wert <- c(6, 7, 6, 5, 5, 6, 4, 5, 7, 8, 5, 8)
daten <- data.frame(gruppe=factor(gruppe), wert)

summary(aov(wert ~ gruppe, daten))
##              Df Sum Sq Mean Sq F value Pr(>F)
## gruppe        2      8   4.000     3.6  0.071 .
## Residuals     9     10   1.111
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

7.6.3.1 Permutationstest zur Varianzanalyse

```
aov_permute <- function(grp, val, B=499) {
  grp <- factor(grp)
  n <- length(val)
  obs <- anova(lm(val ~ grp))$F[1]
  res <- numeric(B)
  for (i in 1:B) {
    index <- sample(n); val.perm <- val[index]
    res[i] <- anova(lm(val.perm ~ grp))$F[1] }
  p <- (sum(res > obs) + 1) / (B+1)
  cat("ANOVA-Permutationstest P=",p,"\n")
}

aov_permute(gruppe, wert)
## ANOVA-Permutationstest P= 0.062
```

7.6.3.2 Stichprobenumfänge und Power

```
npwr.ANOVA <- function(effect, groups, alpha=0.05, power=NULL, n=NULL) {
  if (sum(sapply(list(n, power), is.null)) != 1)
    stop("Fehler: Nur eins von n oder power darf NULL sein")
  f.distr <- quote({
    lambda <- effect^2 * n * groups
    q.alpha <- qf(alpha, groups-1, (n-1)*groups, lower.tail=FALSE)
    pf(q.alpha, groups-1, (n-1)*groups, lambda, lower.tail=FALSE) })
  if (is.null(n)) {
    n <- uniroot(function(n) eval(f.distr)-power, c(2, 1e+05))$root }
  if (is.null(power)) power <- eval(f.distr)
  cat("Stichprobenumfang n =",round(n, 0),"\n",
      "Anzahl der Gruppen =",groups,"\n",
      "Effekt f =",effect,"\n",
      "Signifikanzniveau =",alpha,"\n",
      "Power =",round(power*100,2),"%")
}

npwr.ANOVA(effect=0.353, groups=5, alpha=0.05, power=0.80)
## Stichprobenumfang n = 20
## Anzahl der Gruppen = 5
## Effekt f = 0.353
## Signifikanzniveau = 0.05
## Power = 80 %
```

7.6.4 Multiple Vergleiche

7.6.4.1 Multiple Vergleich nach Tukey-Kramer

Obere Schranken der SR-Verteilung:

```
p <- 0.05
k <- 2:12
v <- c(2:40, 50, 60, 70, 80, 90, 100, 10000)

tab <- matrix(data = NA, nrow = 46, ncol = 11,
              byrow = FALSE, dimnames = NULL)
for (i in 1:46) tab[i,] <- round(qtukey(0.95, k, v[i]), 2)
tab <- cbind(v, tab)
colnames(tab) <- c("v", 2:12)
as.data.frame(tab[1:10,])
```

v	2	3	4	5	6	7	8	9	10	11	12
2	6.08	8.33	9.80	10.88	11.73	12.43	13.03	13.54	13.99	14.40	14.76
3	4.50	5.91	6.82	7.50	8.04	8.48	8.85	9.18	9.46	9.72	9.95
4	3.93	5.04	5.76	6.29	6.71	7.05	7.35	7.60	7.83	8.03	8.21
5	3.64	4.60	5.22	5.67	6.03	6.33	6.58	6.80	6.99	7.17	7.32
6	3.46	4.34	4.90	5.30	5.63	5.90	6.12	6.32	6.49	6.65	6.79
7	3.34	4.16	4.68	5.06	5.36	5.61	5.82	6.00	6.16	6.30	6.43
8	3.26	4.04	4.53	4.89	5.17	5.40	5.60	5.77	5.92	6.05	6.18
9	3.20	3.95	4.41	4.76	5.02	5.24	5.43	5.59	5.74	5.87	5.98
10	3.15	3.88	4.33	4.65	4.91	5.12	5.30	5.46	5.60	5.72	5.83
11	3.11	3.82	4.26	4.57	4.82	5.03	5.20	5.35	5.49	5.61	5.71

Beispiel Antibiotika elementar:

```
A <- c(27, 27, 25, 26, 25); nA <- length(A)
B <- c(26, 25, 26, 25, 24); nB <- length(B)
C <- c(21, 21, 20, 20, 22); nC <- length(C)
grp <- c(rep("A", nA), rep("B", nB), rep("C", nC))
d <- data.frame(Gruppe = factor(grp), Wert = c(A, B, C))

f <- nA + nB + nC - 3
mA <- mean(A); mB <- mean(B); mC <- mean(C)
s <- sqrt((sum((A-mA)^2)+sum((B-mB)^2)+sum((C-mC)^2)) / f)

T.AB <- (mA - mB) / (s*sqrt(0.5*(1/nA + 1/nB))); T.AB
## [1] 2
T.AC <- (mA - mC) / (s*sqrt(0.5*(1/nA + 1/nC))); T.AC
## [1] 13
T.BC <- (mB - mC) / (s*sqrt(0.5*(1/nB + 1/nC))); T.BC
## [1] 11

q <- qtkey(0.95, 3, f); q
## [1] 3.772929
```

Beispiel Antibiotika mit Funktion aov() und TukeyHSD():

```

A <- c(27, 27, 25, 26, 25); nA <- length(A)
B <- c(26, 25, 26, 25, 24); nB <- length(B)
C <- c(21, 21, 20, 20, 22); nC <- length(C)
grp <- c(rep("A", nA), rep("B", nB), rep("C", nC))
d <- data.frame(Gruppe = factor(grp), Wert = c(A, B, C))

model <- aov(Wert ~ Gruppe, data = d);
TukeyHSD(model)
##      Tukey multiple comparisons of means
##      95% family-wise confidence level
##
## Fit: aov(formula = Wert ~ Gruppe, data = d)
##
## $Gruppe
##      diff      lwr      upr      p adj
## B-A -0.8 -2.309172  0.7091716 0.3647720
## C-A -5.2 -6.709172 -3.6908284 0.0000025
## C-B -4.4 -5.909172 -2.8908284 0.0000139

```

Beispiel Antibiotika:

Funktion glht() in library(multcomp)

```

library(multcomp)
grp <- c(rep("A", nA), rep("B", nB), rep("C", nC))
d <- data.frame(Gruppe = factor(grp), Wert = c(A, B, C))

model <- aov(Wert ~ Gruppe, data = d);
summary(glht(model, linfct = mcp(Gruppe = "Tukey"))) # summary(model)
##
##      Simultaneous Tests for General Linear Hypotheses
##
## Multiple Comparisons of Means: Tukey Contrasts
##
##
## Fit: aov(formula = Wert ~ Gruppe, data = d)
##
## Linear Hypotheses:
##      Estimate Std. Error t value Pr(>|t|)
## B - A == 0   -0.8000      0.5657  -1.414    0.365
## C - A == 0   -5.2000      0.5657  -9.192 <0.001 ***
## C - B == 0   -4.4000      0.5657  -7.778 <0.001 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## (Adjusted p values reported -- single-step method)

```

Funktion confint() in library(multcomp)

```

confint(glht(model, linfct = mcp(Gruppe = "Tukey")))
##
##      Simultaneous Confidence Intervals
##

```

```

## Multiple Comparisons of Means: Tukey Contrasts
##
##
## Fit: aov(formula = Wert ~ Gruppe, data = d)
##
## Quantile = 2.6684
## 95% family-wise confidence level
##
##
## Linear Hypotheses:
##           Estimate lwr      upr
## B - A == 0 -0.8000 -2.3095  0.7095
## C - A == 0 -5.2000 -6.7095 -3.6905
## C - B == 0 -4.4000 -5.9095 -2.8905

mA-mB + qtkey(0.95, 3, f)*s*sqrt(0.5*(1/nA + 1/nB))      # Berechnung direkt ...
## [1] 2.309172

mA-mB - qtkey(0.95, 3, f)*s*sqrt(0.5*(1/nA + 1/nB))
## [1] -0.7091716

```

7.6.4.3 Multiple Vergleich nach Dunnett

Beispiel Blutzellen:

```
Kontrolle <- c(7.40, 8.50, 7.20, 8.24, 9.84, 8.32)
Praep.A   <- c(9.76, 8.80, 7.68, 9.36)
Praep.B   <- c(12.80, 9.68, 12.16, 9.20, 10.55)

n0 <- length(Kontrolle); nA <- length(Praep.A); nB <- length(Praep.B)
f  <- n0+nA+nB-(3+1)
m0 <- mean(Kontrolle);  mA <- mean(Praep.A);  mB <- mean(Praep.B)
s  <- sqrt((sum((Kontrolle-m0)^2)+sum((Praep.A-mA)^2)+sum((Praep.B-mB)^2)) / f)

D.A <- (mA - m0) / (s*sqrt(1/nA + 1/n0)); D.A
## [1] 0.8205458
D.B <- (mB - m0) / (s*sqrt(1/nB + 1/n0)); D.B
## [1] 3.536499

R    <- sqrt(nA/(n0+nA)) * sqrt(nB/(n0+nB))
cR   <- matrix(c(1, R, R, 1), nrow=2); round(cR,2)
##      [,1] [,2]
## [1,] 1.00 0.43
## [2,] 0.43 1.00

library(mvtnorm)                                # Funktion qmult() in library(mvtnorm)
qmult(0.95, tail = "both.tail", df = f, corr = cR)$quantile
## [1] 2.543653
```

Funktion contrMat() in library(multcomp)

```
library(multcomp)
grp <- c(rep("grp.0", n0), rep("grp.1", nA), rep("grp.2", nB))
d   <- data.frame(Gruppe = factor(grp), Wert = c(Kontrolle, Praep.A, Praep.B))

n   <- c(n0, nA, nB); names(n) <- paste("Gruppe", c("K", "A", "B"), sep=".")
K   <- contrMat(n, type="Dunnett", base=1); K
##
##   Multiple Comparisons of Means: Dunnett Contrasts
##
##               Gruppe.K Gruppe.A Gruppe.B
## Gruppe.A - Gruppe.K      -1         1         0
## Gruppe.B - Gruppe.K      -1         0         1

model <- aov(Wert ~ Gruppe, data = d)
summary(glht(model, linfct = mcp(Gruppe = K), alternative = "greater"))
##
##   Simultaneous Tests for General Linear Hypotheses
##
## Multiple Comparisons of Means: User-defined Contrasts
##
##
## Fit: aov(formula = Wert ~ Gruppe, data = d)
##
```

```
## Linear Hypotheses:
##               Estimate Std. Error t value Pr(>t)
## Gruppe.A - Gruppe.K <= 0   0.6500     0.7584   0.857 0.32498
## Gruppe.B - Gruppe.K <= 0   2.6280     0.7115   3.694 0.00291 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## (Adjusted p values reported -- single-step method)
```

Konfidenzintervalle:

```
confint(glht(model, linfct = mcp(Gruppe = K), alternative = "two.sided"))
##
## Simultaneous Confidence Intervals
##
## Multiple Comparisons of Means: User-defined Contrasts
##
##
## Fit: aov(formula = Wert ~ Gruppe, data = d)
##
## Quantile = 2.5136
## 95% family-wise confidence level
##
##
## Linear Hypotheses:
##               Estimate lwr      upr
## Gruppe.A - Gruppe.K == 0  0.6500 -1.2564  2.5564
## Gruppe.B - Gruppe.K == 0  2.6280  0.8396  4.4164
```

7.6.4.4 Auswahl des Besten nach HSU bei gleichen Stichprobenumfängen

Obere Schranken der Dunett-Verteilung (Tabelle): Längere Laufzeit....

Funktion qDunnett() in library(mvtnorm)

```
library(mvtnorm) # Quantile zur Dunnett-Verteilung
qDunnett <- function(p, df, k, tail="both.tails") {
  m <- k-1
  R <- matrix(0.5, m, m) # Korrelationsmatrix bei gleichen
  for (i in 1:m) R[i, i] <- 1 # Stichprobenumfängen
  temp <- qmvt(p, interval=c(0, 7), tail=tail, df=df, corr=R)[1]
  return(temp$quantile)
}

fg <- c(5,7,10,12,14,16,20,24,28,30,40,50,60,80,120,5000)
rc <- length(fg)
kn <- 2:8
cc <- length(kn)

# tab <- matrix(rep(0, rc*cc), nrow=rc, dimnames=list(fg,kn))
# for (i in 1:rc) { # alpha = 0.10 zweiseitig
#   for (j in 1:cc) tab[i,j] <- round(qDunnett(0.90, df=fg[i], k=kn[j]), 3) }
# tab <- cbind(fg, tab); colnames(tab) <- c("FG", 2:8)
# as.data.frame(tab[1:5,])
#
# tab <- matrix(rep(0, rc*cc), nrow=rc, dimnames=list(fg,kn))
# for (i in 1:rc) { # alpha = 0.05 zweiseitig
#   for (j in 1:cc) tab[i,j] <- round(qDunnett(0.95, df=fg[i], k=kn[j]), 3) }
# tab <- cbind(fg, tab); colnames(tab) <- c("FG", 2:8)
# as.data.frame(tab[1:5,])
```

Beispiel Insektenfallen:

```
insects <- matrix(c(45, 59, 48, 46, 38, 47,
                   21, 12, 14, 17, 13, 17,
                   37, 32, 15, 25, 39, 41,
                   16, 11, 20, 21, 14, 7), byrow=F, nrow=6,
                 dimnames=list(1:6,c("gelb","weiss","rot","blau")))
d <- data.frame(insects); attach(d)
as.data.frame(d)
```

gelb	weiss	rot	blau
45	21	37	16
59	12	32	11
48	14	15	20
46	17	25	21
38	13	39	14
47	17	41	7

```

gefangen <- c(gelb, weiss, rot, blau)
farbe <- as.factor(c(rep("gelb",6),rep("weiss",6),rep("rot",6),rep("blau",6)))
anova <- summary(aov(gefangen ~ farbe)); anova
##           Df Sum Sq Mean Sq F value    Pr(>F)
## farbe      3   4218    1406    30.55 1.15e-07 ***
## Residuals 20    920     46
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
MSE <- anova[[1]][2,3]; MSE
## [1] 46.025

mi <- apply(insects, 2, mean); mi                                     # Mittelwerte nach den Farben
##      gelb      weiss      rot      blau
## 47.16667 15.66667 31.50000 14.83333
n <- nrow(insects); k <- ncol(insects)
MSE <- 0                                                             # berechne MSE (error)
for (i in 1:k) {for (j in 1:n) MSE <- MSE + (insects[j,i]-mi[i])^2}
MSE <- MSE/(k*(n-1))

##### qd <- qDunnett(0.95, df=k*(n-1), k, tail="lower.tail") #####
qd <- 2.191
d <- qd * sqrt(2*MSE/n)                                             # Distanz mit Dunnett-Quantil

for (i in 1:k) {                                                    # Beschränkte Konfidenzintervalle
  m <- mi[i]; mmax <- max(mi[-i])
  lower.ci <- m - mmax - d; lower.cic <- min(lower.ci, 0)
  upper.ci <- m - mmax + d; upper.cic <- max(upper.ci, 0)
  cat("\n\t", round(lower.cic, 3), "\t - \t", round(upper.cic, 3),
      "für", colnames(insects)[i]) }
##
## 0 - 24.248 für gelb
## -40.082 - 0 für weiss
## -24.248 - 0 für rot
## -40.915 - 0 für blau

```

7.6.4.6 Maximum-Modulus-Ansatz

Studentisierte Maximum-Modulus-Verteilung (SMM): längere Laufzeit....

Funktion qmvt() und qmvnorm() in library(mvtnorm)

```
# library(mvtnorm)
# fg <- c(5,10,15,20,25,30,40,50,75,100,200,1000); l <- length(fg)
# k <- c(1:10, 15, 20); m <- length(k)
#
# tab1 <- matrix(rep(NA, l * m), nrow=l, byrow=T)
# alpha <- 0.05
# for (i in 1:(l-1)) { # multivariate t-Verteilung
#   for (j in 1:m) {
#     tab1[i, j] <- qmvt(1-alpha, df = fg[i], tail = "both",
#                       corr=diag(k[j]))$quantile } }
# for (j in 1:m) { # multivariate Normalvert.
#   tab1[l, j] <- qmvnorm(1-alpha, mean=rep(0,k[j]), tail = "both",
#                       sigma=diag(k[j]))$quantile }
# tab1 <- cbind(fg, tab1); row <- c(NA, round(k, 0));
# tab1 <- rbind(row, tab1)
# tab1
#
# tab2 <- matrix(rep(NA, l * m), nrow=l, byrow=T)
# alpha <- 0.01
# for (i in 1:(l-1)) { # multivariate t-Verteilung
#   for (j in 1:m) {
#     tab2[i, j] <- qmvt(1-alpha, df = fg[i], tail = "both",
#                       corr=diag(k[j]))$quantile } }
# for (j in 1:m) { # multivariate Normalvert.
#   tab2[l, j] <- qmvnorm(1-alpha, mean=rep(0,k[j]), tail = "both",
#                       sigma=diag(k[j]))$quantile }
# tab2 <- cbind(fg, tab2); row <- c(NA, round(k, 0))
# tab2 <- rbind(row, tab2)
# tab2
```

Beispiel Arbeitsunfähigkeit:

```
grp <- c("A","B","C","D","E","F")
n.i <- c( 74, 13, 15, 47, 23, 28); N <- sum(n.i)
m.i <- c(14.30, 13.65, 19.57, 16.91, 13.38, 15.89)

mean <- sum(n.i*m.i)/N; s <- 15.26; mean; s
## [1] 15.38315
## [1] 15.26

d.i <- round(m.i - mean, 2); sd.i <- round(sqrt(s/n.i * (N-n.i)/N), 2)
quot <- round(abs(d.i) / sd.i, 2)
SMM <- rep(NA, 6); k <- rep(NA, 6)

tab <- as.data.frame(cbind(grp, n.i, m.i, d.i, sd.i, quot, SMM, k))
names(tab) <- c("Gruppe", "Anzahl", "Mittelwert", "Differenz",
               "Stdabw", "Quotient", "SMM", "k")
```

```
I <- order(tab$Quotient, decreasing=T); tab <- tab[I, ]      # sortieren

SMM <- c(2.66, 2.59, 2.51, 2.40, 2.25, 1.97)                # k=6,...,1 | FG=n-6=194
tab[,8] <- 6:1; tab[,7] <- SMM; tab
```

	Gruppe	Anzahl	Mittelwert	Differenz	Stdabw	Quotient	SMM	k
3	C	15	19.57	4.19	0.97	4.32	2.66	6
4	D	47	16.91	1.53	0.5	3.06	2.59	5
1	A	74	14.3	-1.08	0.36	3	2.51	4
5	E	23	13.38	-2	0.77	2.6	2.40	3
2	B	13	13.65	-1.73	1.05	1.65	2.25	2
6	F	28	15.89	0.51	0.68	0.75	1.97	1

7.6.4.7 Lineare Kontraste nach Scheffe

```
x <- c( 4,  8, 11, 14, 10,  9, 11,  6); mean(x)
## [1] 9.125

y <- c(17, 10, 11, 13, 14,  9, 11, 12, 12,  8); mean(y)
## [1] 11.7

z <- c(12, 16, 11, 12, 17, 22, 12, 16, 17, 13, 19, 12); mean(z)
## [1] 14.91667

grp    <- c(rep(1,8), rep(2,10), rep(3,12))
wert    <- c(x, y, z)
daten   <- data.frame(grp=factor(grp), wert)
aov.mod <- aov(wert ~ grp, daten); summary(aov.mod)
##              Df Sum Sq Mean Sq F value    Pr(>F)
## grp              2  166.4   83.20   8.644 0.00125 **
## Residuals       27  259.9    9.63
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

se.contrast(aov.mod, list(grp=="1", grp=="2", grp=="3"), coef=c(-1, 0, 1))
## [1] 1.416099
```

Funktion `fit.contrast()` aus `library(gmodels)`

```
library(gmodels)
fit.contrast(aov.mod, grp, c(-1, 0, 1))
##              Estimate Std. Error t value    Pr(>|t|)
## grp c( -1 0 1 ) 5.791667   1.416099  4.089874 0.0003487793
## attr(,"class")
## [1] "fit_contrast"
```

7.6.5 H-Test von Kruskal und Wallis

Kritische Schranken mit Funktion `qKruskalWallis()` aus `library(SuppDists)`:

```
library(SuppDists)
k <- c(3,4,5,6)
n <- c(3:10, 12, 14, 16, 18, 20, 25, 30, 40, 50, 1000)
alpha <- c(0.10, 0.05, 0.01)

matrix(data = NA, nrow = 1, ncol = 1, byrow = FALSE, dimnames = NULL)
##      [,1]
## [1,]    NA
tab <- matrix(NA, nrow = 20, ncol = 14, byrow = TRUE)
tab[1,] <- c(NA, rep(0.10, 4), rep(0.05, 4), rep(0.01, 4), NA)
tab[2,] <- c(NA, rep(c(3,4,5,6), 3), NA)

for (i in 3:20) {
  tab[i,] <- c(n[i-2],
    round(qKruskalWallis(0.90, k, k*n[i-2], k*(1/n[i-2])), 3),
    round(qKruskalWallis(0.95, k, k*n[i-2], k*(1/n[i-2])), 3),
    round(qKruskalWallis(0.99, k, k*n[i-2], k*(1/n[i-2])), 3), n[i-2]) }
as.data.frame(tab[3:10, 1:6])
```

V1	V2	V3	V4	V5	V6
3	4.472	5.912	7.286	8.618	5.292
4	4.543	6.039	7.452	8.815	5.546
5	4.569	6.098	7.535	8.917	5.668
6	4.581	6.132	7.585	8.980	5.738
7	4.588	6.154	7.618	9.022	5.783
8	4.593	6.169	7.641	9.052	5.815
9	4.595	6.180	7.659	9.075	5.838
10	4.597	6.189	7.672	9.092	5.856

Beispiel mit Funktion `kruskal.test()`:

```
A <- c(12.1, 14.8, 15.3, 11.4, 10.8)
B <- c(18.3, 49.6, 10.1, 35.6, 26.2, 8.9)
C <- c(12.7, 25.1, 47.0, 16.3, 30.4)
D <- c( 7.3,  1.9,  5.8, 10.1,  9.4)

x <- c(A, B, C, D)
g <- factor(rep(1:4, c(5, 6, 5, 5)), labels = c("A", "B", "C", "D"))

kruskal.test(x, g)
##
##  Kruskal-Wallis rank sum test
##
## data:  x and g
## Kruskal-Wallis chi-squared = 11.53, df = 3, p-value = 0.009179
```

7.6.5.1 Multiple paarweise Vergleiche mittlerer Ränge

Beispiel mit Funktion `kruskalmc()` aus `library(pgirmess)`:

```
library(pgirmess)
demo <- data.frame(y = c(28, 30, 33, 35, 38, 41,
                        36, 39, 40, 43, 45, 50,
                        44, 45, 47, 49, 53, 54),
                  gruppe = factor(c(rep("A",6),rep("B",6),rep("C",6))))

kruskalmc(y ~ gruppe, data=demo)
## Multiple comparison test after Kruskal-Wallis
## p.value: 0.05
## Comparisons
##      obs.dif critical.dif difference
## A-B  5.583333    7.378741    FALSE
## A-C 10.416667    7.378741    TRUE
## B-C  4.833333    7.378741    FALSE
```

Beispiel mit Funktion `oneway__test()` aus `library(coin)`:

```
library(coin)
demo <- data.frame(y = c(28, 30, 33, 35, 38, 41,
                        36, 39, 40, 43, 45, 50,
                        44, 45, 47, 49, 53, 54),
                  gruppe = factor(c(rep("A",6),rep("B",6),rep("C",6))))

Nemenyi <- oneway__test(y ~ gruppe, data = demo,
                       ytrafo = function(data) trafo(data, numeric_trafo = rank),
                       xtrafo = function(data) trafo(data, factor_trafo = function(x)
model.matrix(~x - 1) %*% t(contrMat(table(x), "Tukey"))),
                       teststat = "max")

Nemenyi
##
## Asymptotic K-Sample Fisher-Pitman Permutation Test
##
## data: y by gruppe (A, B, C)
## chi-squared = 11.453, df = 2, p-value = 0.003258

drop(pvalue(Nemenyi, method = "single-step"))
## [1] 0.003257908
```

Nemenyi-Damico-Wolfe-Dunn Test - Funktion `independence__test()` aus `library(coin)`:

```
library(coin)
demo <- data.frame(y = c(28, 30, 33, 35, 38, 41,
                        36, 39, 40, 43, 45, 50,
                        44, 45, 47, 49, 53, 54),
                  gruppe = factor(c(rep("A",6),rep("B",6),rep("C",6))))

kw <- kruskal__test(y ~ gruppe, data = demo,
                   distribution = approximate(nresample=9999)); kw
```

```
##
## Approximative Kruskal-Wallis Test
##
## data: y by gruppe (A, B, C)
## chi-squared = 11.453, p-value = 4e-04

it <- independence_test(y ~ gruppe, data = demo,
                        distribution = approximate(nresample = 50000),
                        ytrafo = function(data) trafo(data, numeric_trafo = rank_trafo),
                        xtrafo = function(data) trafo(data, factor_trafo = function(x)
                        model.matrix(~x - 1) %*% t(contrMat(table(x), "Tukey")))); it

##
## Approximative General Independence Test
##
## data: y by gruppe (A, B, C)
## maxT = 3.3814, p-value = 0.00028
## alternative hypothesis: two.sided

pvalue(it, method = "single-step")
##
## B - A 0.17276
## C - A 0.00028
## C - B 0.27752
```

7.6.5.4 Trendtest nach Jonckheere

```
jonckheere.test<-function(x, g) { # Jonckheere-Terpstra Test
  x <- table(g,x); nco <- ncol(x); nro <- nrow(x); summe <- 0
  for(j in 1:(nco - 1))
    for(i in 1:(nro - 1))
      summe <- summe + x[i, j] * (0.5 * sum(x[(i + 1):nro, j]) +
                                sum(x[(i + 1):nro, (j + 1):nco]))
  for(k in 1:(nro - 1))
    summe <- summe + x[k, nco] * 0.5 * sum(x[(k + 1):nro, nco])
  n <- sum(x); nip <- apply(x, 1, sum); npj <- apply(x, 2, sum)
  expect <- (n^2 - sum(nip^2))/4
  u1 <- n * (n - 1) * (2 * n + 5) - sum(nip * (nip - 1) * (2 * nip + 5)) -
      sum(npj * (npj - 1) * (2 * npj + 5))
  u2 <- sum(nip * (nip - 1) * (nip - 2)) * sum(npj * (npj - 1) * (npj - 2))
  u3 <- sum(nip * (nip - 1)) * sum(npj * (npj - 1))
  v <- u1/72 + u2/(36 * n * (n - 1) * (n - 2)) + u3/(8 * n * (n - 1))
  zval <- (summe - expect)/sqrt(v); pval <- 1 - pnorm(zval)
  cat("Jonckheere-Terpstra Test : ", " Statistik =",
      round(zval, 3), " P =", round(pval, 3), "\n")
}
```

Beispiel 1):

```
A <- c(30, 31, 34, 34, 37, 39)
B <- c(36, 38, 41, 41, 45, 48)
C <- c(44, 45, 47, 49, 50, 50)
```

```
werte <- c(A, B, C); n <- c(6, 6, 6)
grp <- as.ordered(factor(rep(1:length(n),n)))

jonckheere.test(werte, grp)
## Jonckheere-Terpstra Test : Statistik = 3.768 P = 0
```

Beispiel 2):

```
A <- c(106, 114, 116, 127, 145)
B <- c(110, 125, 143, 148, 151)
C <- c(136, 139, 149, 160, 174)
werte <- c(A, B, C); n <- c(5, 5, 5)

grp <- as.ordered(factor(rep(1:length(n),n)))

jonckheere.test(werte, grp)
## Jonckheere-Terpstra Test : Statistik = 2.272 P = 0.012
```

Beispiel 2) mit Funktion oneway_test() aus library(coin):

```
A <- c(106, 114, 116, 127, 145)
B <- c(110, 125, 143, 148, 151)
C <- c(136, 139, 149, 160, 174)
werte <- c(A, B, C); n <- c(5, 5, 5)

library(coin)
d <- data.frame(werte=werte, grp=factor(c(rep("A",5),rep("B",5),rep("C",5))))

oneway_test(werte ~ grp, data=d, scores = list(grp = c(1, 2, 3)))
##
## Asymptotic Linear-by-Linear Association Test
##
## data: werte by grp (A < B < C)
## Z = 2.4246, p-value = 0.01533
## alternative hypothesis: two.sided
```

7.6.6 Varianzanalyse für Messwiederholungen

Beispiel Gewichtsreduktion:

```
diet <- data.frame(effect = c(1.5, 1.4, 1.4, 1.2, 1.4,
                             2.7, 2.9, 2.1, 3.0, 3.3,
                             2.1, 2.2, 2.4, 2.0, 2.5,
                             1.3, 1.0, 1.1, 1.3, 1.5),
                  patient = factor(paste("pat", rep(1:5, 4), sep="")),
                  zeit     = factor(paste("T", rep(c(1, 2, 3, 4),
                                                  c(5, 5, 5, 5)), sep="")),
                  row.names = NULL)

summary(aov(effect ~ zeit + Error(patient), data=diet))
##
## Error: patient
##           Df Sum Sq Mean Sq F value Pr(>F)
## Residuals  4  0.393  0.09825
##
## Error: Within
##           Df Sum Sq Mean Sq F value    Pr(>F)
## zeit       3  8.154   2.7178   41.87 1.24e-06 ***
## Residuals 12  0.779   0.0649
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

7.6.7 Friedman-Test

Tabelle mit Funktion qFriedman() aus library(SuppDists):

```
library(SuppDists)
k <- c(3,4,5,6)
n <- c(3:10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 45,
      50, 60, 70, 80, 90, 100, 1000)

alpha <- c(0.05, 0.01)

tab <- matrix(NA, nrow = 27, ncol = 10, byrow = TRUE)
tab[1,] <- c(NA, rep(0.05,4), rep(0.01, 4), NA)
tab[2,] <- c(NA, rep(c(3,4,5,6), 2), NA)

for (i in 3:27) {
  tab[i,] <- c(n[i-2], round(qFriedman(0.95, k, n[i-2]), 3),
               round(qFriedman(0.99, k, n[i-2]), 3), n[i-2]) }

colnames(tab) <- c("n",3:6,3:6,"n")
as.data.frame(tab[13:20,])
```

n	3	4	5	6	3	4	5	6	n
16	6.125	7.800	9.350	10.893	9.125	10.875	12.725	14.500	16
18	5.778	7.800	9.378	10.921	8.778	10.933	12.800	14.571	18
20	6.100	7.800	9.400	10.943	9.100	10.980	12.840	14.629	20
25	6.000	7.776	9.408	10.971	8.880	11.016	12.928	14.720	25
30	6.067	7.800	9.413	10.990	8.867	11.120	12.973	14.781	30
35	6.000	7.783	9.440	10.980	9.029	11.143	13.029	14.824	35
40	6.000	7.800	9.440	11.000	9.075	11.160	13.060	14.857	40
45	6.000	7.787	9.440	11.010	9.111	11.173	13.084	14.883	45

Beispiel Schokoladensorten:

```
y <- matrix(c( 2.2, 2.0, 1.8,
               2.4, 1.8, 1.6,
               2.5, 1.9, 1.7,
               1.7, 2.5, 1.9),
            nrow = 4, byrow = TRUE,
            dimnames = list(Person = as.character(1:4),
                           Sorte = LETTERS[1:3]))
as.data.frame(y)
```

A	B	C
2.2	2.0	1.8
2.4	1.8	1.6
2.5	1.9	1.7
1.7	2.5	1.9

```
n <- dim(y)[1];    k <- dim(y)[2]
R <- matrix(rep(NA, n*k), nrow=n, byrow=TRUE)           # Rangzahlen
for (i in 1:n) R[i,] <- rank(-y[i,]); R
##      [,1] [,2] [,3]
## [1,]    1    2    3
## [2,]    1    2    3
## [3,]    1    2    3
## [4,]    3    1    2
Ri2 <- colSums(R)^2; Ri2
## [1]  36  49 121

stat <- (12/(n*k*(k+1)))*sum(Ri2) - 3*n*(k+1); stat
## [1] 3.5
pval <- 1 - pchisq(stat, k-1); pval
## [1] 0.1737739

friedman.test(y)                                         # Funktion friedman.test()
##
## Friedman rank sum test
##
## data:  y
## Friedman chi-squared = 3.5, df = 2, p-value = 0.1738
```

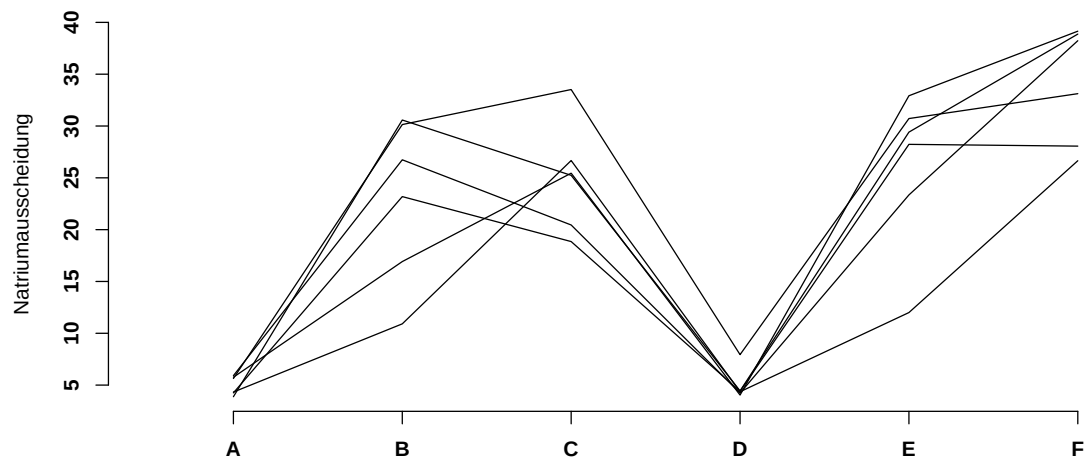
7.6.7.2 Multiple paarweise Vergleiche nach Wilcoxon und Wilcox

Beispiel Diuretika:

```
diuret <- data.frame(block = factor(rep(1:6, rep(6,6))),  
  diuretikum = factor(rep(c("A","B","C","D", "E","F"), 6)),  
  natrium = c(3.88, 30.58, 25.24, 4.44, 29.41, 38.87,  
    5.64, 30.14, 33.52, 7.94, 30.72, 33.12,  
    5.76, 16.92, 25.45, 4.04, 32.92, 39.15,  
    4.25, 23.19, 18.85, 4.40, 28.23, 28.06,  
    5.91, 26.74, 20.45, 4.23, 23.35, 38.23,  
    4.33, 10.91, 26.67, 4.36, 12.00, 26.65))  
  
reshape(diuret, timevar="diuretikum", idvar="block", direction="wide")
```

	block	natrium.A	natrium.B	natrium.C	natrium.D	natrium.E	natrium.F
1	1	3.88	30.58	25.24	4.44	29.41	38.87
7	2	5.64	30.14	33.52	7.94	30.72	33.12
13	3	5.76	16.92	25.45	4.04	32.92	39.15
19	4	4.25	23.19	18.85	4.40	28.23	28.06
25	5	5.91	26.74	20.45	4.23	23.35	38.23
31	6	4.33	10.91	26.67	4.36	12.00	26.65

```
par(mfcol=c(1,1), lwd=3, font.axis=2, bty="l", ps=12)  
matplot(t(matrix(diuret$natrium, ncol = 6, byrow = TRUE)),  
  type = "l", col = 1, lty = 1, axes = FALSE,  
  ylab = "Natriumausscheidung", xlim = c(0.5, 6.5))  
axis(1, at = 1:6, labels = levels(diuret$diuretikum)); axis(2)
```



Beispiel Diuretika:

Funktion `symmetry_test()` in `library(coin)`

```
library(coin) # multiple paarweise Vergleiche
friedman_test(natrium ~ diuretikum | block, data = diuret)
##
## Asymptotic Friedman Test
##
## data: natrium by
## diuretikum (A, B, C, D, E, F)
## stratified by block
## chi-squared = 23.333, df = 5, p-value = 0.0002915
library(multcomp)
friedm <- symmetry_test(natrium ~ diuretikum | block, data = diuret,
  xtrafo = function(data) trafo(data, factor_trafo = function(x)
    model.matrix(~ x - 1) %*% t(contrMat(table(x), "Tukey"))),
  ytrafo = function(data) trafo(data, numeric_trafo = rank,
    block = diuret$block),
  teststat = "max",
)
pvalue(friedm) # Friedman-Test
## [1] 0.001588278
## 99 percent confidence interval:
## 0.001464350 0.001712206

drop(round(pvalue(friedm, method = "single-step"), 5))
## B - A C - A D - A E - A F - A C - B D - B E - B F - B D - C
## 0.18815 0.09154 0.99963 0.03960 0.00158 0.99963 0.33868 0.98982 0.63622 0.18815
## E - C F - C E - D F - D F - E
## 0.99963 0.81999 0.09154 0.00507 0.93999
```

7.6.7.3 Page-Test für geordnete Alternativen

Funktion `page.trend.test()` in `library(crank)`

```
library(crank)
Gutachter <- matrix(c(2,2,1,2,2,1,3,1,1,      # Objekt B
                     1,3,2,3,1,2,2,2,4,      # Objekt C
                     4,1,3,1,4,3,1,4,2,      # Objekt D
                     3,4,4,4,3,4,4,3,3),      # Objekt A
                    nrow=9,byrow=F)

Page <- page.trend.test(Gutachter, ranks=TRUE)
Page
##
## Page test for ordered alternatives
## L = 252  p(table) <=.01
```

Geordnete Alternativen mit der Funktion `friedman_test()` in `library(coin)`:

```
g <- data.frame(block = factor(rep(1:9, rep(4,9))),
                objekt = ordered(rep(c("Obj1", "Obj2", "Obj3", "Obj4"), 9)),
                note = c(2,1,4,3,      2,3,1,4,      1,2,3,4,
                        2,3,1,4,      2,1,4,3,      1,2,3,4,
                        3,2,1,4,      1,2,4,3,      1,4,2,3))

library(coin)
friedman_test(note ~ objekt | block, data = g)
##
## Asymptotic Page Test
##
## data:  note by
##  objekt (Obj1 < Obj2 < Obj3 < Obj4)
##  stratified by block
## Z = 3.1177, p-value = 0.001823
## alternative hypothesis: two.sided
```

7.6.7.4 Spannweitenrangtest nach Quade

Beispiel Marktanalyse:

```
y <- matrix(c( 5, 4, 7, 10, 12, 1, 3, 1, 0, 2,
              16, 12, 22, 22, 35, 5, 4, 3, 5, 4,
              10, 9, 7, 13, 10, 19, 18, 28, 37, 58,
              10, 7, 6, 8, 7),
            nrow = 7, byrow = TRUE,
            dimnames = list(Store = as.character(1:7),
                           Brand = LETTERS[1:5]))
as.data.frame(y)
```

	A	B	C	D	E
1	5	4	7	10	12
2	1	3	1	0	2
3	16	12	22	22	35
4	5	4	3	5	4
5	10	9	7	13	10
6	19	18	28	37	58
7	10	7	6	8	7

```
k <- dim(y)[1];    b <- dim(y)[2]
R <- matrix(rep(NA,k*b), nrow=k, byrow=TRUE)      # Rangzahlen
for (i in 1:k) R[i,] <- rank(y[i,])
as.data.frame(R)
```

	V1	V2	V3	V4	V5
1	2.0	1.0	3.0	4.0	5.0
2	2.5	5.0	2.5	1.0	4.0
3	2.0	1.0	3.5	3.5	5.0
4	4.5	2.5	1.0	4.5	2.5
5	3.5	2.0	1.0	5.0	3.5
6	2.0	1.0	3.0	4.0	5.0
7	5.0	2.5	1.0	4.0	2.5

```
range <- rep(NA, k)                                     # Spannweiten
for (i in 1:k) range[i] <- max(y[i,]) - min(y[i,])
Qi <- rank(range)

S <- Qi * (R - (b + 1)/2)                               # Scores
A2 <- sum(S^2)                                           # Q-gesamt
B <- sum(colSums(S)^2)/k                                # Q-zwischen
stat <- (k-1)*B / (A2-B); stat                          # Teststatistik
## [1] 3.829252
pval <- 1 - pf(stat, b-1, (b-1)*(k-1)); pval
## [1] 0.01518902
```

```
quade.test(y)                                # quade.test() in library(stats)
##
## Quade test
##
## data:  y
## Quade F = 3.8293, num df = 4, denom df = 24, p-value = 0.01519
```

Multiple paarweise Vergleiche mit Funktion `posthoc.quade.test()` in `library(PMCMR)`

```
library(PMCMR)
posthoc.quade.test(y, dist="TDist", p.adj="none")
##
## Pairwise comparisons using posthoc-Quade test with TDist approximation
##
## data:  y
##
##      A      B      C      D
## B 0.2087 -      -      -
## C 0.8401 0.2874 -      -
## D 0.1477 0.0102 0.1021 -
## E 0.0416 0.0021 0.0269 0.5172
##
## P value adjustment method: none
```

7.6.8 Zweifache Varianzanalyse

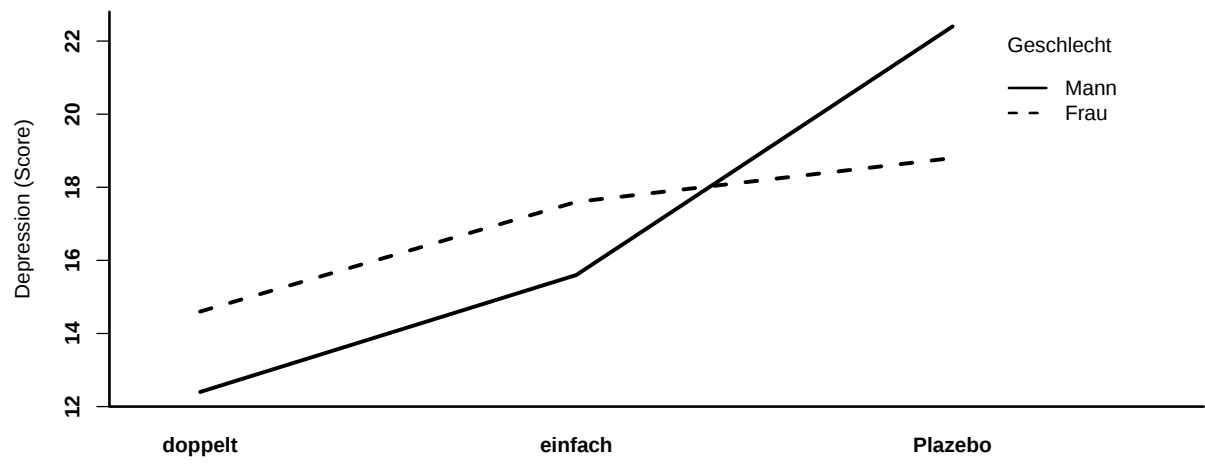
Beispiel Antidepressiva:

```
depr <- data.frame(
  score = c(22, 25, 22, 21, 22, 16, 16, 16, 15, 15, 13, 12, 12, 13, 12,
            18, 19, 17, 21, 19, 19, 20, 17, 16, 16, 16, 14, 16, 13, 14),
  geschl = factor(c(rep("Mann", 15), rep("Frau", 15))),
  therap = factor(rep(c(rep("Plazebo", 5), rep("einfach", 5),
                        rep("doppelt", 5)), 2)))
as.data.frame(depr[1:5,])
```

score	geschl	therap
22	Mann	Plazebo
25	Mann	Plazebo
22	Mann	Plazebo
21	Mann	Plazebo
22	Mann	Plazebo

```
summary(aov(score ~ therap + geschl + geschl:therap, depr))
##              Df Sum Sq Mean Sq F value    Pr(>F)
## therap         2  253.4   126.7    74.529 5.06e-11 ***
## geschl         1    0.3     0.3     0.176   0.678
## therap:geschl   2   54.2    27.1   15.941 3.94e-05 ***
## Residuals     24   40.8     1.7
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

par(mfrow=c(1,1), lwd=2, font.axis=2, bty="l", ps=12)
interaction.plot(depr$therap, depr$geschl, depr$score, main=" ",
  trace.label="Geschlecht", lwd=3, xlab=" ", ylab="Depression (Score)")
```

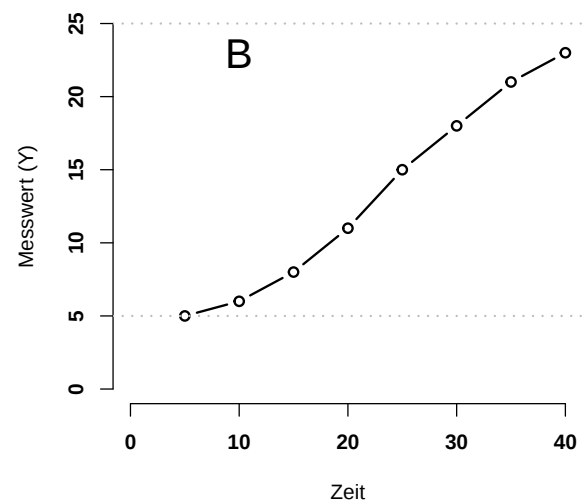
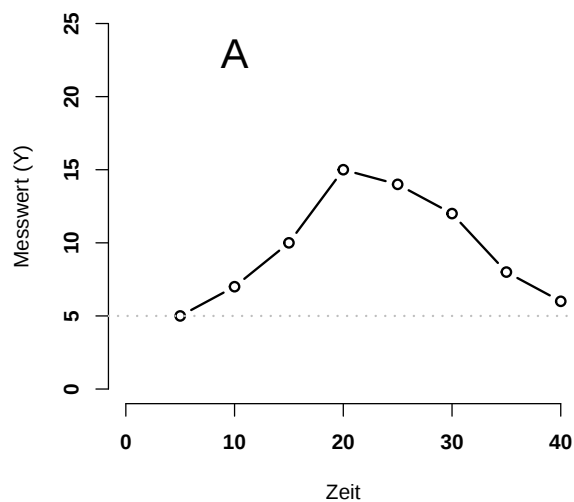


7.6.9 Analyse von wiederholten Messungen

Zeitlicher Verlauf:

```
Zeit <- c(5, 10, 15, 20, 25, 30, 35, 40)
y1 <- c(5, 7, 10, 15, 14, 12, 8, 6)
y2 <- c(5, 6, 8, 11, 15, 18, 21, 23)

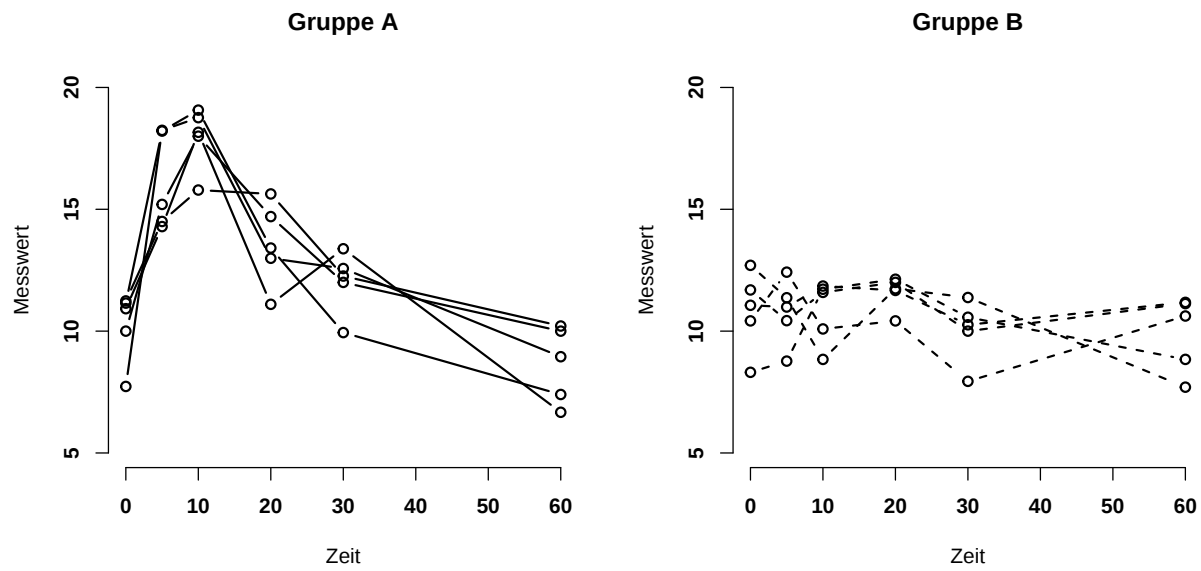
par(mfrow=c(1,2), lwd=1.8, font.axis=2, bty="n", ps=12)
plot(Zeit, y1, type="b", ylab="Messwert (Y)", ylim=c(0,25), xlim=c(0,40))
abline(h=5, lty=3, col="grey")
text(10, 23, "A", cex=2)
plot(Zeit, y2, type="b", ylab="Messwert (Y)", ylim=c(0,25), xlim=c(0,40))
text(10, 23, "B", cex=2)
abline(h=5, lty=3, col="grey")
abline(h=25, lty=3, col="grey")
```



Beispiel:

```
daten <- read.csv2("repeated.csv")
attach(daten)
Zeit <- c(0, 5, 10, 20, 30, 60)

par(mfrow=c(1,2), lwd=1.7, font.axis=2, bty="n", ps=12)
plot(Zeit, daten[1,3:8], type="b", ylab="Messwert", main="Gruppe A",
     ylim=c(5,20))
for (i in 2:5) lines(Zeit, daten[i, 3:8], type="b")
plot(Zeit, daten[6,3:8], type="b", ylab="Messwert", main="Gruppe B",
     ylim=c(5,20), lty=2)
for (i in 7:10) lines(Zeit, daten[i, 3:8], type="b", lty=2)
```



```

AUC  <- function(y, t, n) {                                     # Area Under Curve
  F <- rep(NA, (n-1))
  for (i in 1:(n-1)) F[i] <- (t[i+1]-t[i])*(y[i]+y[i+1])
  sum(F)/2
}

REGR <- function(y, t, n) {                                     # Regressions-Koeffizient
  sum((y-mean(y))*(t-mean(t)))/sum((t-mean(t))^2)
}

daten <- read.csv2("repeated.csv")
attach(daten)
Zeit <- c(0, 5, 10, 20, 30, 60)

daten$Max  <- apply(daten[,3:8], 1, max, na.rm = T)             # Tabelle zum Beispiel
daten$AUC  <- apply(daten[,3:8], 1, AUC, t=Zeit, n=6)           # Maximum
daten$REGR <- apply(daten[,5:8], 1, REGR, t=Zeit[3:6], n=4)     # AUC
daten                                           # Regression

```

Gruppe	Prob	t0	t5	t10	t20	t30	t60	Max	AUC	REGR
A	1	10.00	15.20	18.00	14.70	12.00	10.00	18.00	773.000	-0.1478571
A	2	10.92	14.29	18.16	11.10	13.38	6.67	18.16	713.600	-0.1957857
A	3	7.73	18.24	18.76	12.99	12.57	8.95	18.76	766.775	-0.1690000
A	4	11.15	18.21	19.07	13.41	9.94	7.40	19.07	705.850	-0.2096429
A	5	11.24	14.50	15.79	15.63	12.26	10.21	15.79	773.675	-0.1184286
B	6	11.69	10.43	11.59	12.00	10.00	11.13	12.00	655.250	-0.0127857
B	7	12.70	11.37	8.84	11.74	11.38	7.70	12.70	615.400	-0.0451429
B	8	8.31	8.77	11.72	12.13	10.57	8.84	12.13	617.825	-0.0646429
B	9	10.42	12.42	10.09	10.42	7.94	10.62	12.42	586.125	0.0090000
B	10	11.06	10.99	11.85	11.67	10.26	11.18	11.85	661.075	-0.0130714

```

t1 <- t.test(Max ~ Gruppe, data = daten)           # Maximum
max <- c(t1$estimate,t1$statistic,t1$p.value)
t2 <- t.test(AUC ~ Gruppe, data = daten)          # AUC
auc <- c(t2$estimate,t2$statistic,t2$p.value)
t3 <- t.test(REGR ~ Gruppe, data = daten)         # Aenderung
reg <- c(t3$estimate,t3$statistic,t3$p.value)
t4 <- t.test(t60 ~ Gruppe, data = daten)          # letzter Wert
lst <- c(t4$estimate,t4$statistic,t4$p.value)
tab <- as.data.frame(rbind(max, auc, reg, lst),
                     row.names = c("Max", "AUC", "REGR", "t60"))
colnames(tab) <- c("mean A", "mean B", "t-stat", "p-val")
as.data.frame(tab)

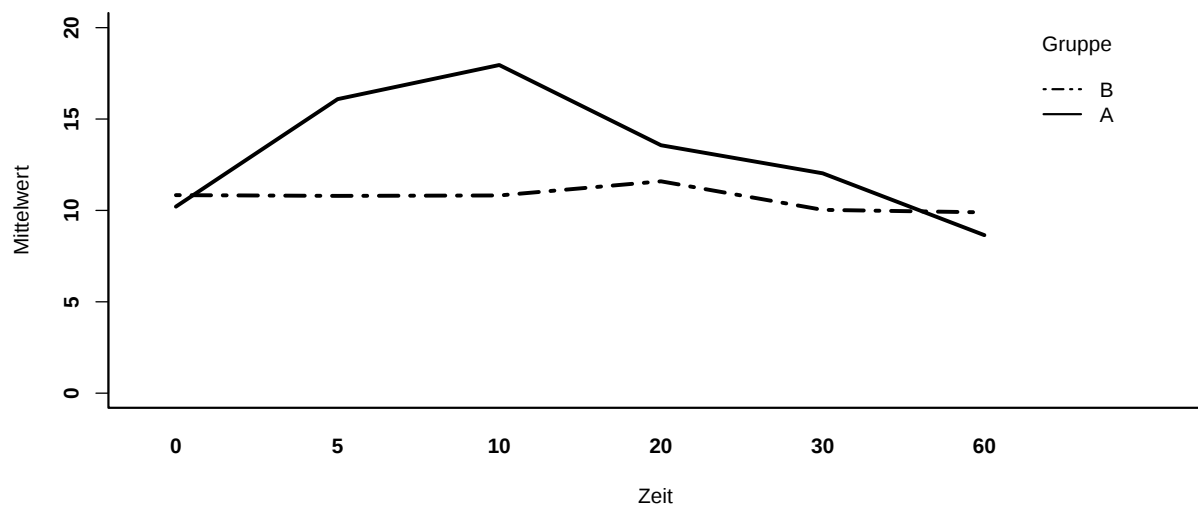
```

	mean A	mean B	t-stat	p-val
Max	17.9560000	12.2200000	9.636377	0.0003354
AUC	746.5800000	627.1350000	5.816149	0.0004093
REGR	-0.1681429	-0.0253286	-6.810586	0.0001701
t60	8.6460000	9.8940000	-1.265131	0.2414327

7.6.9.2 ANOVA für wiederholte Messungen (gemischte Modelle)

```
daten <- read.csv2("repeated.csv")
Zeit <- c(0, 5, 10, 20, 30, 60)
neu <- reshape(daten, varying=list(c("t0", "t5", "t10", "t20", "t30", "t60")),
               v.names=c("Wert"), timevar="Zeit",
               times=c(0, 5, 10, 20, 30, 60), idvar="Prob", direction="long")

par(mfrow=c(1,1), lwd=1.7, font.axis=2, bty="l", ps=12)
interaction.plot(neu$Zeit, factor(neu$Gruppe), neu$Wert, lty=c(1,12), lwd=3,
                 ylim=c(0,20), ylab="Mittelwert", xlab="Zeit", trace.label="Gruppe")
```



```
rep.aov <- aov(neu$Wert ~ factor(neu$Gruppe)*factor(neu$Zeit) + Error(factor(neu$Prob)))
summary(rep.aov)
##
## Error: factor(neu$Prob)
##               Df Sum Sq Mean Sq F value    Pr(>F)
## factor(neu$Gruppe)  1  87.94   87.94    76.69 2.26e-05 ***
## Residuals           8   9.17    1.15
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Error: Within
##               Df Sum Sq Mean Sq F value    Pr(>F)
## factor(neu$Zeit)  5 185.92   37.18   15.97 1.22e-08 ***
## factor(neu$Gruppe):factor(neu$Zeit)  5 134.07   26.81   11.52 6.28e-07 ***
## Residuals       40  93.11    2.33
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

7.6.10 Versuchsplanung

Beispiel Feldversuch:

```
Amm <- c(rep(0,4),rep(1,4),rep(0,4),rep(1,4),rep(0,4),rep(1,4),rep(0,4),rep(1,4))
Magn <- c(rep(0,8),rep(1,8),rep(0,8),rep(1,8))
Mist <- c(rep(0,16),rep(1,16))
yield <- c(19.2,15.5,17.0,11.7,20.6,16.9,19.5,21.9,18.9,20.2,16.7,
           20.7,25.3,27.6,29.1,25.4,20.8,18.5,20.1,19.2,26.8,17.8,
           18.6,19.0,22.2,18.6,22.3,21.1,27.7,28.6,28.7,28.5)
data <- data.frame(block=gl(8,4), Amm=factor(Amm),
                   Magn=factor(Magn), Mist=factor(Mist), yield=yield)

yield.aov1 <- aov(yield ~ block, data)
summary(yield.aov1)
##              Df Sum Sq Mean Sq F value    Pr(>F)
## block          7  484.2   69.17   12.92 8.91e-07 ***
## Residuals     24  128.5    5.35
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

yield.aov2 <- aov(yield ~ Amm*Magn*Mist, data)
summary(yield.aov2)
##              Df Sum Sq Mean Sq F value    Pr(>F)
## Amm           1  196.52   196.52   36.700 2.95e-06 ***
## Magn          1  192.57   192.57   35.963 3.43e-06 ***
## Mist          1   32.60    32.60    6.089 0.02112 *
## Amm:Magn       1   52.79    52.79    9.858 0.00444 **
## Amm:Mist       1    5.70     5.70    1.064 0.31267
## Magn:Mist      1    0.69     0.69    0.129 0.72270
## Amm:Magn:Mist  1    3.32     3.32    0.619 0.43907
## Residuals     24  128.51    5.35
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

7.7 Die Analyse von Häufigkeiten

7.7.1 Vergleich relativer Häufigkeiten

Beispiel Präsidentschaftswahlen (Bootstrap)

```
n.obama <- 120; n.mccain <- 80; n.other <- 10
data <- c(rep("Obama",n.obama),
          rep("McCain", n.mccain), rep("andere",n.other))

bootstrap.stat <- function(data) {                                # Bootstrap
  b.smpl <- sample(data, length(data), replace=TRUE)
  (sum(b.smpl=="Obama") - sum(b.smpl=="McCain"))/length(data)
}

p.distr <- replicate(500, bootstrap.stat(data))
round(quantile(p.distr, probs=c(0.025, 0.25, 0.50, 0.75, 0.975)), 4)
## 2.5% 25% 50% 75% 97.5%
## 0.0619 0.1429 0.1905 0.2333 0.3143
```

Beispiel Biodiversitäten (Shannon-Index):

```
food      <- c("Eiche","Mais","Brombeere","Buche","Kirsche","Sonstige")
michigan  <- c(47, 35, 7, 5, 3, 2)
louisiana <- c(48, 23, 11, 13, 8, 2)
jay.diet  <- as.data.frame(cbind(food, michigan, louisiana))
jay.diet
```

food	michigan	louisiana
Eiche	47	48
Mais	35	23
Brombeere	7	11
Buche	5	13
Kirsche	3	8
Sonstige	2	2

```
shannon.index <- function(x) {
  n <- sum(x); k <- length(x)
  (n*log10(n) - sum(x*log10(x)))/n
}

H.1 <- shannon.index(michigan); H.1
## [1] 0.5403328
H.2 <- shannon.index(louisiana); H.2
## [1] 0.6327828
```

```
shannon.test <- function(x, y, alpha) {
  sign <- 1 - alpha/2
  nx <- sum(x); kx <- length(x)
```

```

ny <- sum(y); kx <- length(y)
Hx <- (nx*log10(nx) - sum(x*log10(x)))/nx           # Shannon-Index 1
Hy <- (ny*log10(ny) - sum(y*log10(y)))/ny           # Shannon Index 2
s2x <- (sum(x*log10(x)^2) - (sum(x*log10(x)))^2/nx)/nx^2
s2y <- (sum(y*log10(y)^2) - (sum(y*log10(y)))^2/ny)/ny^2
stat <- abs((Hx - Hy) / sqrt(s2x + s2y))             # Teststatistik
fg <- round(nx*ny*(s2x+s2y)^2 / (ny*s2x^2 + nx*s2y^2), 0)
p.val <- (1 - pt(stat, fg))*2                        # P-Wert zweiseitig
cat(" Shannon-Wiener-Index in Population X =", round(Hx, 4),
    "und in Population y =", round(Hy, 4),"\n",
    "Tesstatistik:", round(stat, 4),
    "t-Verteilung mit", fg, "Freiheitsgraden:",
    round(qt(sign, fg), 4), "- Pwert", round(p.val, 4),"\n")
}

shannon.test(michigan, louisiana, alpha=0.05)
## Shannon-Wiener-Index in Population X = 0.5403 und in Population y = 0.6328
## Tesstatistik: 1.909 t-Verteilung mit 196 Freiheitsgraden: 1.9721 - Pwert 0.0577

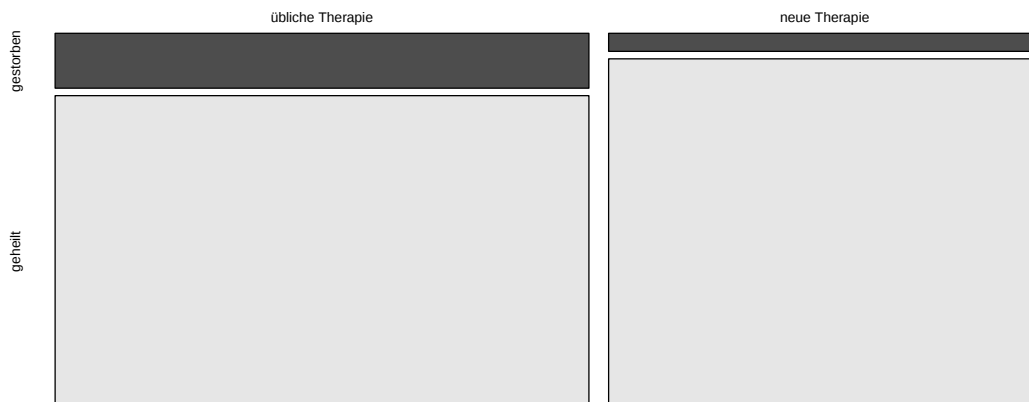
```

7.7.2 Analyse von Vierfeldertafeln

```
tab <- matrix(c(15, 85, 4, 77), nrow=2, ncol=2, byrow=TRUE)
dimnames(tab) <- list(c("übliche Therapie", "neue Therapie"),
                      c("gestorben", "geheilt"))
as.data.frame(tab)
```

	gestorben	geheilt
übliche Therapie	15	85
neue Therapie	4	77

```
mosaicplot(tab, col=TRUE, main=" ")
```



```
chisq.test(tab, correct=FALSE) # ohne Korrektur
##
## Pearson's Chi-squared test
##
## data:  tab
## X-squared = 4.8221, df = 1, p-value = 0.0281

chisq.test(tab, correct=TRUE) # mit Korrektur
##
## Pearson's Chi-squared test with Yates' continuity correction
##
## data:  tab
## X-squared = 3.8107, df = 1, p-value = 0.05093
```

7.7.2.1 Fallzahl und Power zum Vierfeldertest

```
z.alpha <- qnorm(0.975); z.beta <- qnorm(0.90)

p1 <- 0.38;      q1 <- 1 - p1
p2 <- 0.30;      q2 <- 1 - p2
p <- (p1 + p2)/2; q <- 1 - p
n <- (z.alpha * sqrt(2*p*q) + z.beta *
      sqrt(p1*q1+p2*q2))^2 / ((p2 - p1)^2); n
## [1] 734.7537
```

```
power.prop.test(p1=0.3, p2=0.38, sig.level =0.05, power = 0.90)
##
##      Two-sample comparison of proportions power calculation
##
##              n = 734.7537
##              p1 = 0.3
##              p2 = 0.38
##      sig.level = 0.05
##              power = 0.9
##      alternative = two.sided
##
## NOTE: n is number in *each* group

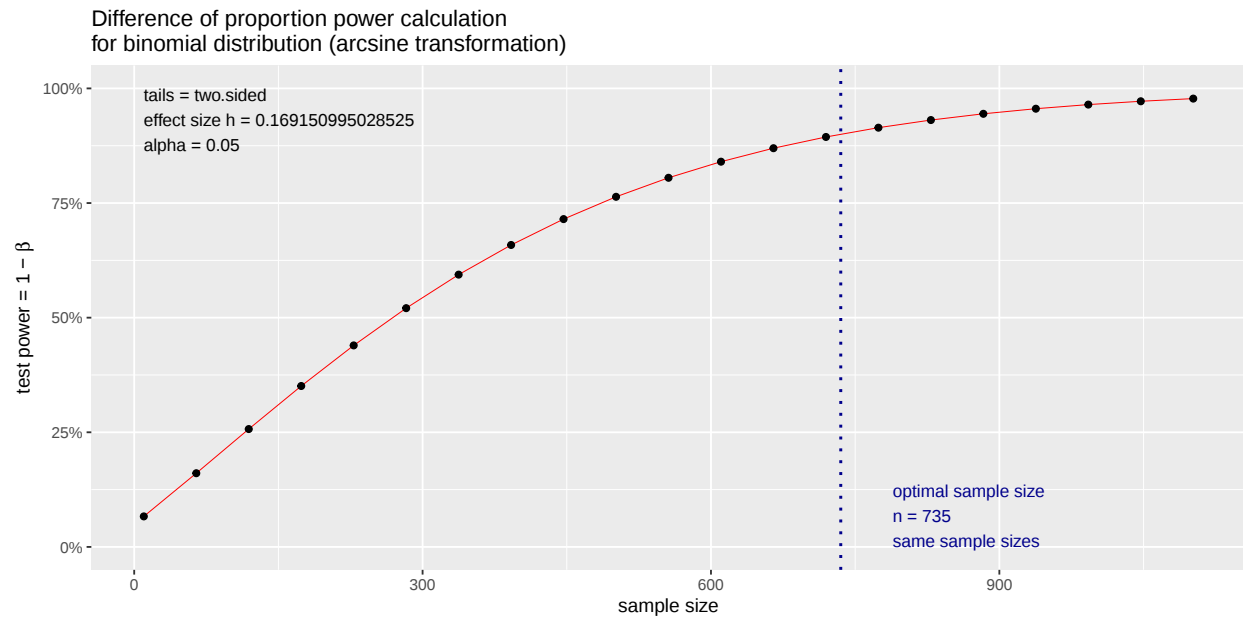
power.prop.test(p1=0.3, p2=0.38, sig.level =0.05, n=735)
##
##      Two-sample comparison of proportions power calculation
##
##              n = 735
##              p1 = 0.3
##              p2 = 0.38
##      sig.level = 0.05
##      power = 0.9000955
##      alternative = two.sided
##
## NOTE: n is number in *each* group
```

Funktion ES.h() und pwr.2p.test() in library(pwr)

```
library(pwr)
effect <- ES.h(0.38, 0.30); effect
## [1] 0.169151
p2p <- pwr.2p.test(h = effect, sig.level = 0.05, power = 0.90);
p2p
##
##      Difference of proportion power calculation for binomial distribution
##
##              h = 0.169151
##              n = 734.4749
##      sig.level = 0.05
##      power = 0.9
##      alternative = two.sided
```

```
##  
## NOTE: same sample sizes
```

```
plot(p2p)
```



7.7.3 Spezielle Risiko- und Effektmaße

Relatives Risiko und Odds Ratio

```
a <- 24; b <- 96; c <- 48; d <- 592
tab <- matrix(c(a, b, c, d), nrow=2, ncol=2, byrow=TRUE)
dimnames(tab) <- list(c("exponiert", "nicht exponiert"),
                      c("krank", "nichtkrank"))
as.data.frame(tab)
```

	krank	nichtkrank
exponiert	24	96
nicht exponiert	48	592

```
IR.exp <- a / (a+b); IR.exp           # Inzidenzrate exponiert
## [1] 0.2

IR.nexp <- c / (c+d); IR.nexp         # Inzidenzrate nicht exponiert
## [1] 0.075

delta <- IR.exp - IR.nexp; delta      # zuschreibbares Risiko
## [1] 0.125

psi <- IR.exp / IR.nexp; psi          # relatives Risiko
## [1] 2.666667

omega <- (a*d) / (b*c); omega         # Odds Ratio
## [1] 3.083333
```

Funktion `oddsratio()` in `library(vcd)`

```
library(vcd)
library(Hmisc)
a <- 24; b <- 96; c <- 48; d <- 592
tab <- matrix(c(a, b, c, d), nrow=2, ncol=2, byrow=TRUE)
dimnames(tab) <- list(c("exponiert", "nicht exponiert"),
                      c("krank", "nichtkrank")); tab

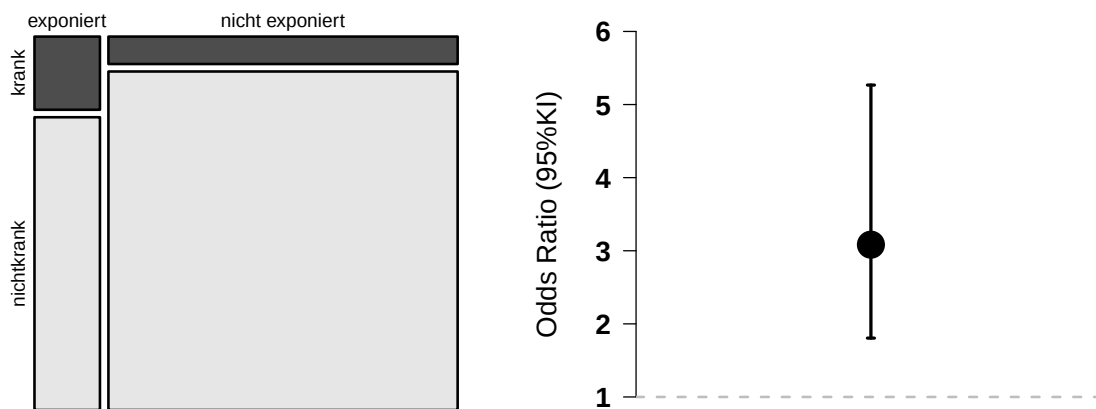
##                krank nichtkrank
## exponiert         24         96
## nicht exponiert   48        592
OR <- loddsratio(tab, log=FALSE)

sor <- summary(OR); sor           # OR Schätzung
##
## z test of coefficients:
##
##                Estimate Std. Error z value
## exponiert:nicht exponiert/krank:nichtkrank 3.08333 0.84218 3.6611
##                Pr(>|z|)
## exponiert:nicht exponiert/krank:nichtkrank 0.0002511 ***
## ---
```

```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

cor <- confint(OR); cor                                     # Konfidenzintervall
##                                                         2.5 %    97.5 %
## exponiert:nicht exponiert/krank:nichtkrank 1.805189 5.266454

par(mfrow=c(1,2), lwd=2, font.axis=2.5, bty="n", ps=16)
mosaicplot(tab, col=TRUE, main=" ")
u <- cor[1]; v <- cor[1]; o <- cor[2]
errbar( x=1, v, o, u , xlim=c(0.9, 1.1), ylim=c(1,6), las=1, xaxt="n",
        cex=3, lwd=2.5, xlab=" ", ylab="Odds Ratio (95%KI)")
abline(h=1, lty=2, col="grey")
```



7.7.3.3 Stichprobenumfänge

Fall-Kontroll-Studien mit mehreren Kontrollen:

```
smpl.matched.cc <- function(alpha, beta, MM, OR, ff) {
  zalpha <- qnorm(alpha/2, lower.tail = FALSE)
  zbeta <- qnorm(beta, lower.tail=FALSE)
  ss <- (OR-1)*ff*(1-ff)/((1-ff)+ ff*OR)
  nn <- (MM+1)*((zalpha*(1+OR) + 2*zbeta*sqrt(OR))^2)/(2*MM*(OR^2 - 1)*ss)
  round(nn, 0)
}

smpl.matched.cc(alpha=0.05, beta=0.10, 1:4, OR=1.5, ff=0.1)
## [1] 1206 905 804 754

result <- matrix(NA, nrow=6, ncol=5)
or <- c(1.5,2,2.5,3,3.5,4)
for (i in 1:length(or))
  result[i, ] <- c(or[i], smpl.matched.cc(0.05, 0.10, 1:4, or[i], 0.1))
```

```
colnames(result) <- c("OR", "1:1", "1:2", "1:3", "1:4")
as.data.frame(result)
```

OR	1:1	1:2	1:3	1:4
1.5	1206	905	804	754
2.0	368	276	245	230
2.5	193	145	129	121
3.0	126	94	84	79
3.5	92	69	61	57
4.0	72	54	48	45

Hypothesentest in Kohortenstudien:

```
npwr.RR <- function(P2, RR, N=NULL, alpha=0.05, power=NULL, einseitig=TRUE, r=1) {
  if (sum(sapply(list(N, power), is.null)) != 1)
    stop("Entweder 'N' oder die 'Power' müssen NULL gesetzt werden!")
  if (einseitig) zalph <- qnorm(1-alpha) else zalph <- qnorm(1-alpha/2)
  p.c <- (P2*(r*RR+1))/(r+1)
  if (is.null(N)) {
    N <- (r+1)/(r*(RR-1)^2*P2^2 * (zalph*sqrt((r+1)*p.c*(1-p.c))
      + qnorm(power) * sqrt(RR*P2*(1-RR*P2)+r*P2*(1-P2)))^2 }
  if (is.null(power)) {
    zpower <- (P2*abs(RR-1)*sqrt(N*r) - zalph*(r+1)*sqrt(p.c*(1-p.c))) /
      sqrt((r+1)*(RR*P2*(1-RR*P2)+r*P2*(1-P2)))
    power <- pnorm(zpower) }
  cat("Stichprobenumfang und Power zum relativen Risiko","\n",
      "Stichprobenumfang N:",round(N, 0)," \n",
      "Power:",round(power,5)," \n")
}

npwr.RR(P2=0.2, RR=2.0, alpha=0.05, einseitig=TRUE, power=0.90)
## Stichprobenumfang und Power zum relativen Risiko
## Stichprobenumfang N: 176
## Power: 0.9

npwr.RR(P2=0.2, RR=2.0, alpha=0.05, einseitig=TRUE, N=176)
## Stichprobenumfang und Power zum relativen Risiko
## Stichprobenumfang N: 176
## Power: 0.8999
```

7.7.3.4 Der expositionsbedingte Anteil (PAR)

Beispiel Framingham-Studie:

```
a <- 72; b <- 684; c <- 20; d <- 553; N <- a+b+c+d

RR <- ( a*c + a*d ) / ( a*c + b*c ) ; RR
## [1] 2.728571

Pexp <- ( a + b ) / N ; Pexp
```

```
## [1] 0.5688488

PAR <- Pexp*(RR-1) / (1 + Pexp*(RR-1)); PAR
## [1] 0.4957888

var <- ( c*N*( a*d*(N-c )+ b*c ^ 2 ) ) / ( ( a+c ) ^3*( c+d ) ^ 3 )
se <- sqrt(var)
KIu <- PAR - 1.96*se; KIo <- PAR + 1.96*se; KIu; KIo
## [1] 0.3046911
## [1] 0.6868865
```

7.7.4 Exakter Test nach R.A. Fisher

Beispiel Tea Tasting Lady:

```
TeaTasting <- matrix(c(3, 1, 1, 3), nr = 2,
  dimnames = list(Guess = c("Milk", "Tea"), Truth = c("Milk", "Tea")))
TeaTasting
##      Truth
## Guess Milk Tea
## Milk    3   1
## Tea     1   3

fisher.test(TeaTasting, alternative = "greater")      # Funktion fisher.test()
##
## Fisher's Exact Test for Count Data
##
## data:  TeaTasting
## p-value = 0.2429
## alternative hypothesis: true odds ratio is greater than 1
## 95 percent confidence interval:
##  0.3135693      Inf
## sample estimates:
## odds ratio
##  6.408309
```

```
tab <- matrix(c(2, 8, 10, 4), byrow=TRUE, nr = 2); tab
##      [,1] [,2]
## [1,]    2    8
## [2,]   10    4

fisher.test(tab, alternative="less", conf.level=0.95)      # Funktion fisher.test()
##
## Fisher's Exact Test for Count Data
##
## data:  tab
## p-value = 0.01804
## alternative hypothesis: true odds ratio is less than 1
## 95 percent confidence interval:
##  0.0000000 0.6965009
## sample estimates:
## odds ratio
##  0.1121872
```

7.7.5 Äquivalenztest zweier Binomialwahrscheinlichkeiten

Intervallinklusion (TOST):

```
TOST_int <- function(r1, n1, r2, n2, delta, alpha) {  
  level <- (1-2*alpha)*100  
  p1 <- r1/n1; p2 <- r2/n2; z <- qnorm(1-2*alpha)  
  d <- p1*(1-p1)/n1 + p2*(1-p2)/n2 + ((1/n1)+(1/n2))/2  
  lu <- p1 - p2 - z * sqrt(d); lo <- p1 - p2 + z * sqrt(d)  
  cat("Das",level,"%-KI zur Differenz",p1-p2,"lautet",lu,"bis",lo,"\\n",  
      "mit Bezug zum Äquivalenzintervall",-delta,"bis",+delta,"\\n") }
```

Ansatz nach Dunnett und Gent:

```
TOST_test <- function(r1, n1, r2, n2, delta) {  
  p1 <- r1/n1; p2 <- r2/n2  
  p1d <- (r1 + r2 + delta*n2)/(n1+n2); p2d <- p1d - delta  
  z1 <- (p1 - p2 - delta)/sqrt((p1d*(1-p1d)/n1)+(p2d*(1-p2d))/n2)  
  p1d <- (r1 + r2 - delta*n2)/(n1+n2); p2d <- p1d + delta  
  z2 <- (p1 - p2 + delta)/sqrt((p1d*(1-p1d)/n1)+(p2d*(1-p2d))/n2)  
  P <- pnorm(z1) + (1-pnorm(z2))  
  cat("Der P-Wert für den zweiseitigen Test auf Äquivalenz von ",p1,"versus",p2,"\\n",  
      "mit Delta =",delta,"beträgt P=",P,"\\n") }
```

Beispiel Therapievergleich:

```
TOST_int(120, 200, 57, 100, 0.15, 0.05)  
## Das 90 %-KI zur Differenz 0.03 lautet -0.1053297 bis 0.1653297  
## mit Bezug zum Äquivalenzintervall -0.15 bis 0.15  
  
TOST_test(120, 200, 57, 100, 0.15)  
## Der P-Wert für den zweiseitigen Test auf Äquivalenz von 0.6 versus 0.57  
## mit Delta = 0.15 beträgt P= 0.02449961
```

Hinweis zur Studienplanung:

```
alpha <- 0.05; beta <- 0.20; p1 <- 0.58; p2 <- 0.60; delta <- 0.15  
n <- ((qnorm(1-alpha) + qnorm(1-beta))^2 *  
      (p1*(1-p1) + p2*(1-p2))) / (delta-(p1-p2))^2  
ceiling(n)  
## [1] 104
```

7.7.6 McNemar Vorzeichentest

Beispiel Nachweis der Wirksamkeit:

```
wirk <- matrix(c(8, 16, 5, 11), nr=2, byrow=TRUE,
               dimnames = list(verum=c("stark", "schwach"),
                                placebo=c("stark", "schwach")))
as.data.frame(wirk)
```

	stark	schwach
stark	8	16
schwach	5	11

```
mcnemar.test(wirk, correct=TRUE)
##
## McNemar's Chi-squared test with continuity correction
##
## data:  wirk
## McNemar's chi-squared = 4.7619, df = 1, p-value = 0.0291

discordant <- c(wirk[1,2], wirk[2,1])           # diskordante Ergebnisse
nd          <- sum(discordant)
x           <- min(discordant)
pbinom(x, nd, 0.5)                               # Binomialwahrscheinlichkeit (exakt)
## [1] 0.01330185

binom.test(x, nd, p=0.5)                         # Funktion binom.test()
##
## Exact binomial test
##
## data:  x and nd
## number of successes = 5, number of trials = 21, p-value = 0.0266
## alternative hypothesis: true probability of success is not equal to 0.5
## 95 percent confidence interval:
##  0.08217588 0.47165983
## sample estimates:
## probability of success
##          0.2380952
```

Funktion mcnemar() in library(exact2x2)

```
library(exact2x2)
mcnemar.exact(wirk) # Funktion mcnemar.exact()
##
## Exact McNemar test (with central confidence intervals)
##
## data:  wirk
## b = 16, c = 5, p-value = 0.0266
## alternative hypothesis: true odds ratio is not equal to 1
## 95 percent confidence interval:
```

```
## 1.120172 11.169022
## sample estimates:
## odds ratio
## 3.2
```

Beispiel Urlaubsländer (Konfidenzintervall):

```
urlaub <- matrix(c(71, 3, 16, 10), nrow=2, byrow=TRUE,
  dimnames = list(c("B +", "B -"), c("A +", "A -")))
as.data.frame(urlaub)
```

	A +	A -
B +	71	3
B -	16	10

```
mcnemar.test(urlaub)
##
## McNemar's Chi-squared test with continuity correction
##
## data: urlaub
## McNemar's chi-squared = 7.5789, df = 1, p-value = 0.005905
```

Funktion binom.confint() in library(binom)

```
library(binom)
binom.confint(3, 19, method="exact")
```

method	x	n	mean	lower	upper
exact	3	19	0.1578947	0.0338262	0.3957846

Power und Fallzahl zum McNemar-Test:

```
npwr.mcnemar <- function(p, psi, alpha=0.05, n=NULL, power=NULL) {
  if (sum(sapply(list(n, power), is.null)) != 1)
    stop("exactly one of 'n' or 'power' must be NULL")
  zalpha <- qnorm(1-alpha/2)
  if (is.null(n)) {
    zbeta <- qnorm(power)
    n <- (zalpha*sqrt(psi+1) + zbeta*sqrt((psi+1) - (psi-1)^2*p))^2 /
      (p*(psi-1)^2)
  }
  if (is.null(power)) {
    zbeta <- (sqrt(n)*sqrt(p)*(psi-1) - qnorm(1-alpha/2)*sqrt(psi + 1)) /
      sqrt((psi + 1) - p * (psi - 1)^2)
    power <- pnorm(zbeta)
  }
  cat("Stichprobenumfang und Power zum McNemar-Test", "\n",
    "Stichprobenumfang n:", round(n, 0), "\n",
    "Power:", round(power, 5), "\n")
}
```

```

}

npwr.mcnemar(p=0.125, psi=3.2, n=40)
## Stichprobenumfang und Power zum McNemar-Test
## Stichprobenumfang n: 40
## Power: 0.68298

npwr.mcnemar(p=0.125, psi=2.0, power=0.90)
## Stichprobenumfang und Power zum McNemar-Test
## Stichprobenumfang n: 248
## Power: 0.9

```

7.7.7 Test nach Mantel-Haenszel

```
MH.test <- function(tab, conf.level=0.95, correct=T) {
  ##### 3-dim. Tabelle tab[2,2,k]: 1-Exposition / 2-Krankheit / 3-Stratum #####
  zquant <- qnorm(conf.level)                                # Signifikanzniveau
  cval <- ifelse(correct==T, 0.5, 0)                        # Kontinuitätskorrektur (?)
                                                         # Zwischenergebnisse

  si <- dim(tab)[3]; OR <- V <- E <- w <- S <- numeric(si)

  for (i in 1:si) {                                         # OR - Statistik je Stratum
    OR[i] <- (tab[1,1,i]*tab[2,2,i]) / (tab[1,2,i]*tab[2,1,i])
    E[i] <- colSums(tab[,i])[1] * rowSums(tab[,i])[1] / sum(tab[,i])
    V[i] <- (colSums(tab[,i])[1] * colSums(tab[,i])[2] * rowSums(tab[,i])[1]
              * rowSums(tab[,i])[2]) / (sum(tab[,i])^2 * (sum(tab[,i]) - 1))
    w[i] <- (tab[1,2,i] * tab[2,1,i]) / sum(tab[,i]) }

  ORmh <- sum(w*OR)/sum(w)                                # Mantel-Haenszel Odds-Ratio/Test)
  CHImh <- (sum(tab[1,1,]) - sum(E) - cval)^2 / sum(V)
  pval1 <- pchisq(CHImh, df=1, lower.tail=F)
  cio <- ORmh**((1 + 1.96/sqrt(CHImh))); ciu <- ORmh**((1 - 1.96/sqrt(CHImh))
                                                         # Test Effekt-Modifikation (Confounding)

  for (i in 1:si) S[i] <- (tab[1,1,i]*tab[2,2,i] -
                           ORmh*tab[1,2,i]*tab[2,1,i])^2 / (ORmh*V[i]*sum(tab[,i])^2)
  CHIefm <- sum(S); pval2 <- pchisq(CHIefm, df=1, lower.tail=F)

  cat("\n", "*** Mantel-Haenszel Test (Effekt-Modifikation/Confounding) ***", "\n",
      "Odds-Ratio in Strata:", round(OR, 2), "\n",
      "Mantel-Haenszel OR:", round(ORmh, 2), "\n",
      "Chiquadrat-MH:", round(CHImh, 4), "(P = ", round(pval1, 5), ")", "\n",
      "Konfidenzintervall:", round(ciu, 2), "-", round(cio, 2), "\n",
      "Chiquadrat-Effekt:", round(CHIefm, 4), "(P = ", round(pval2, 5), ")", "\n") }
}
```

Beispiel:

```
tab <- array(c(15, 4, 85, 77, 20, 7, 56, 51), dim = c(2, 2, 2),
            dimnames = list( K = c("I", "II"), E = c("+", "-"),
                              Geschl = c("maennl", "weibl")))
as.data.frame(tab)
```

	+ .maennl	- .maennl	+ .weibl	- .weibl
I	15	85	20	56
II	4	77	7	51

```
MH.test(tab, conf.level=0.95, correct=T)
##
## *** Mantel-Haenszel Test (Effekt-Modifikation/Confounding) ***
## Odds-Ratio in Strata: 3.4 2.6
## Mantel-Haenszel OR: 2.91
## Chiquadrat-MH: 7.8977 (P = 0.00495 )
```

```

##      Konfidenzintervall: 1.38 - 6.14
##      Chiquadrat-Effekt: 0.1204 (P = 0.7286 )

mantelhaen.test(tab, correct = T, conf.level = 0.95)
##
##      Mantel-Haenszel chi-squared test with continuity correction
##
## data:  tab
## Mantel-Haenszel X-squared = 7.8977, df = 1, p-value = 0.00495
## alternative hypothesis: true common odds ratio is not equal to 1
## 95 percent confidence interval:
##  1.410224 6.016843
## sample estimates:
## common odds ratio
##      2.912919

```

7.7.7.1 Breslow-Day-Test

```

breslow.day.test <- function(x, OR=NA) {

  if(is.na(OR)) {                                     # OR nach Mantel-Haenszel
    OR = mantelhaen.test(x)$estimate
    names(OR) = " " }
  k <- dim(x)[3]                                     # Schichten
  a <- hat.a <- Var.a <- numeric(k)                  # Zwischenergebnisse
  X2.stat <- 0
  for (j in 1:k) {                                   # Randsummen
    mj <- apply(x[, ,j], MARGIN=1, sum)
    nj <- apply(x[, ,j], MARGIN=2, sum)

    coef <- c(-mj[1]*nj[1]*OR, nj[2]-mj[1]+OR*(nj[1]+mj[1]), 1-OR)
    sols <- Re(polyroot(coef))                       # 0 < hat.aj <= min(n1_j, m1_j)
    hat.aj <- sols[(0 < sols) & (sols <= min(nj[1],mj[1]))]
                                                    # weitere Schätzungen

    hat.bj <- mj[1]-hat.aj
    hat.cj <- nj[1]-hat.aj
    hat.dj <- mj[2]-hat.cj

                                                    # Varianz
    Var.aj <- (1/hat.aj + 1/hat.bj + 1/hat.cj + 1/hat.dj)^(-1)
    aj <- x[1,1,j]                                   # beobachtete Häufigkeiten
                                                    # Berechnung der Teststatistik
    X2.stat <- X2.stat + as.numeric((aj - hat.aj)^2 / Var.aj)
                                                    # Zwischenergebnisse
    a[j] <- aj; hat.a[j] <- hat.aj; Var.a[j] <- Var.aj
  }

                                                    # Korrektur nach Tarrone
  X2.stat <- as.numeric(X2.stat - (sum(a) - sum(hat.a))^2/sum(Var.a))
                                                    # Berechnung des P-Wertes
  p <- 1-pchisq(X2.stat, df=k-1)

  return(unlist(list(OR = round(OR, 3), Statistik = round(X2.stat, 3),
    df = round(k-1, 0), P.Wert = round(p,6))))
}

```

Beispiel Alkoholgenuss:

```

alcohol <- array(c(1,0,9,106,4,5,26,164,25,21,29,138,
  42,34,27,139,19,36,18,88,5,8,0,31),
  dim=c(2,2,6), dimnames=list(c("exponiert","nicht exponiert"),
  c("Fall","Kontrolle"),
  c("25-34","35-44","45-54","55-64","65-74",>74")))

mantelhaen.test(alcohol, alternative="two.sided", correct=TRUE)
##
## Mantel-Haenszel chi-squared test with continuity correction
##
## data: alcohol
## Mantel-Haenszel X-squared = 83.215, df = 1, p-value < 2.2e-16

```

```
## alternative hypothesis: true common odds ratio is not equal to 1
## 95 percent confidence interval:
## 3.562131 7.467743
## sample estimates:
## common odds ratio
## 5.157623

breslow.day.test(alcokol, OR=NA)
##      OR. Statistik      df      P.Wert
## 5.158000 9.299000 5.000000 0.097704

breslow.day.test(alcokol, OR=2) # H0: OR=2
##      OR Statistik      df      P.Wert
## 2.000000 10.783000 5.000000 0.055863
```

7.7.8 Der $k \times 2$ -Felder-Test nach Brandt und Snedecor

Beispiel Therapieerfolg:

```
erfolg <- matrix(c(14, 22, 18, 16, 8, 2), nr=3, byrow=T,
  dimnames = list(heilung=c("geheilt-x", "geheilt-x+y", "gestorben"),
  therapie=c("symptomatisch", "spezifisch")))
as.data.frame(erfolg)
```

	symptomatisch	spezifisch
geheilt-x	14	22
geheilt-x+y	18	16
gestorben	8	2

```
chisq.test(erfolg, correct = TRUE)
##
## Pearson's Chi-squared test
##
## data:  erfolg
## X-squared = 5.4954, df = 2, p-value = 0.06407
```

7.7.8.1 Multipler Vergleich von Anteilen - Marascuilo-Prozedur

Beispiel Haarfarben:

```
haare <- matrix(c(32, 43, 16, 9, 55, 65, 64, 16), nrow=2, byrow=T)
colnames(haare) <- c("schwarz", "braun", "blond", "rot")
as.data.frame(haare)
```

	schwarz	braun	blond	rot
	32	43	16	9
	55	65	64	16

```
chisq.test(haare)
##
## Pearson's Chi-squared test
##
## data:  haare
## X-squared = 8.9872, df = 3, p-value = 0.02946

x <- haare[1,]; n <- colSums(haare); p <- x/n; k <- length(p)

dif <- matrix(rep(NA, 4*((k*(k-1))/2))), nrow=4, byrow=T)
row.names(dif) <- c("i", "j", "Differenz", "krit.Wert")

m <- 1
for (i in 1:(k-1)) {
```

```

for (j in (i+1):k) {
  dif[1, m] <- i; dif[2, m] <- j
  dif[3, m] <- abs(p[i] - p[j])
  dif[4, m] <- sqrt(qchisq(0.95, df=k-1) *
                    (p[i]*(1-p[i])/n[i] + p[j]*(1-p[j])/n[j]))
  m <- m + 1
}
}
dif[1:2,] <- round(dif[1:2,], 0)
dif[3:4,] <- round(dif[3:4,], 4)

colnames(dif) <- c("schwarz-Braun", "schwarz-blond", "schwarz-rot",
                  "braun-blond", "braun-rot", "blond-rot")
as.data.frame(dif)

```

	schwarz-Braun	schwarz-blond	schwarz-rot	braun-blond	braun-rot	blond-rot
i	1.0000	1.0000	1.0000	2.0000	2.0000	3.0000
j	2.0000	3.0000	4.0000	3.0000	4.0000	4.0000
Differenz	0.0303	0.1678	0.0078	0.1981	0.0381	0.1600
krit.Wert	0.1955	0.1911	0.3048	0.1816	0.2989	0.2961

```

pairwise.prop.test(x, n, p.adjust.method="holm")      # Adjustierte P-Werte
##
## Pairwise comparisons using Pairwise comparison of proportions
##
## data:  x out of n
##
##      schwarz braun blond
## braun 1.000   -   -
## blond 0.131  0.037 -
## rot   1.000  1.000 0.682
##
## P value adjustment method: holm

```

7.7.8.3 Power- und Fallzahlabeschätzung

Beispiel fairer Würfel:

```

alpha <- 0.05                                     # Signifikanzniveau
quant <- qchisq(alpha, df=5, lower.tail = FALSE)
n <- 120                                           # Anzahl der Würfe
pi0 <- rep(1/6, 6)                               # Pi unter der Nullhypothese
pi1 <- c(rep(3/20, 5), 1/4)                     # Pi unter der Alternative
effect <- sum((pi1-pi0)^2/pi0)
l <- n * effect; l                               # Nichtzentralitätsparameter
## [1] 6

power <- pchisq(quant, df=5, ncp=l, lower.tail=FALSE) # Power
power
## [1] 0.4328759

```

```

lrange <- seq(6, 20, by=0.01); i <- 0 # Bereich für lambda
while (power < 0.80) {
  i <- i+1; power <- pchisq(quant, df=5, ncp=lrange[i], lower.tail=FALSE) }

round(power, 4) # Power über 80%
## [1] 0.8001

nc <- lrange[i] / effect # Anzahl der Fälle
ceiling(nc)
## [1] 257

```

7.7.9 Cochran-Armitage-Test auf linearen Trend

```
tabtrend <- function(tab, scores, transpose=FALSE) {
  if (any(dim(tab)==2)) {if (transpose==TRUE) {tab <- t(tab)}}
  if (dim(tab)[1]!=2)
    {stop("Cochran-Armitage nur in (2,k)-Tafel", call.=FALSE)}
  nidot <- apply(tab,2,sum); n <- sum(nidot) # Summen und Scores
  scri <- scores; scrq <- sum(scri*nidot)/n
  p.i <- tab[1,] / nidot # beobachtete Anteile
  p <- sum(tab[1,])/n
  chi <- 1/(p*(1-p))*(sum(nidot*((p.i-p)^2))); chi # chi-Quadrat total
  b <- sum(nidot*(p.i-p)*(scri-scrq))/sum(nidot*(scri-scrq)^2)
  pi.h <- p + b*(scri-scrq)
  chi.e <- (1/(p*(1-p)))*sum(nidot*(p.i-pi.h)^2); chi.e # Abweichung
  chi.t <- b^2/(p*(1-p))*sum(nidot*(scri-scrq)^2); chi.t # Trend
  z <- sqrt(chi.t)
  p <- 2*pnorm(abs(z), lower.tail=FALSE) # P-Wert zweiseitig
  cat(name="Cochran-Armitage Test auf Trend","\n",
      "Chi-Quadrat-Trend :", chi.trend=round(chi.t, 3)," p =", p.wert=p, "\n",
      "Chi-Quadrat-Fehler:", chi.err=round(chi.e, 3),"\n",
      "Chi-Quadrat-gesamt:", chi.gesamt=round(chi,3),"\n")
}}
```

Beispiel Alkoholkonsum und Fehlbildungen:

```
malform <- matrix(c(48, 38, 5, 1, 1, 17066, 14464, 788, 126, 37),
  nrow=2, byrow=T,
  dimnames = list(Fehlbildung=c("ja","nein"),
    alkohol=c("0","<1","1-2","3-5",">5")))
as.data.frame(malform)
```

	0	<1	1-2	3-5	>5
ja	48	38	5	1	1
nein	17066	14464	788	126	37

```
tabtrend(malform, c(0,0.5,1.5,4,7), transpose=FALSE)
## Cochran-Armitage Test auf Trend
## Chi-Quadrat-Trend : 6.57 p = 0.01037041
## Chi-Quadrat-Fehler: 5.512
## Chi-Quadrat-gesamt: 12.082
```

Funktion prop.trend.test():

```
x <- malform[1,]; n <- malform[1,] + malform[2,]
prop.trend.test(x, n, c(0, 0.5, 1.5, 4, 7))
##
## Chi-squared Test for Trend in Proportions
##
## data: x out of n ,
## using scores: 0 0.5 1.5 4 7
## X-squared = 6.5701, df = 1, p-value = 0.01037
```

7.7.10 Vergleich mehrerer Anteile mit einem Standard

```
ind <- function(d, dir="greater") { # Indikatorfunktion
  n <- length(d); s.p <- rep(NA, n); s.n <- rep(NA, n)
  for (i in 1:n) {
    if (d[i]>0) s.p[i]<-1 else s.p[i]<- 0
    if (d[i]<0) s.n[i]<-1 else s.n[i]<- 0 }
  if (dir=="greater") result <- s.p else
    if (dir=="less") result <- s.n else NA
  result
}

prop.ref.test <- function(p.0, x, n, alternative="zweiseitig") {
  k <- length(x)
  if (alternative=="zweiseitig") { # zweiseitig
    B <- sum((x - n*p.0)^2/(n*p.0*(1-p.0)))
    p.val <- pchisq(B, df=k, lower.tail = FALSE)
    cat("B (zweiseitig) =",B," ( P =",round(p.val,8),")\n")
  } else if (alternative=="größer") { # einseitig 'größer'
    diff <- x - n*p.0
    B.plus <- sum((diff*ind(diff, "greater"))^2/(n*p.0*(1-p.0)))
    theta <- pbinom(round(n*p.0), n, prob=p.0, lower.tail = FALSE)
    pv1 <- dbinom(1:k, k, prob=theta)
    pv2 <- pchisq(B.plus, df=1:k, lower.tail = FALSE)
    p.val <- sum(pv1*pv2)
    cat("B (einseitig größer) =",B.plus," ( P =",round(p.val,8),")\n")
  } else if (alternative=="kleiner") { # einseitig 'kleiner'
    diff <- x - n*p.0
    B.min <- sum((diff*ind(diff, "less"))^2/(n*p.0*(1-p.0)))
    theta <- pbinom(round(n*p.0), n, prob=p.0, lower.tail = TRUE)
    pv1 <- dbinom(1:k, k, prob=theta)
    pv2 <- pchisq(B.min, df=1:k, lower.tail = FALSE)
    p.val <- sum(pv1*pv2)
    cat("B (einseitig kleiner) =",B.min," ( P =",round(p.val,8),")\n")
  }
}
```

Beispiel mit drei Anteilen zweiseitig:

```
p.0 <- 0.60
x <- c(59, 107, 48); n <- c(81, 151, 114)

prop.ref.test(p.0, x, n, alternative="zweiseitig")
## B (zweiseitig) = 28.19595 , ( P = 3.3e-06 )
```

Beispiel mit zwei Anteilen größer::

```
p.0 <- 0.60
x <- c(107, 48); n <- c(151, 114)

prop.ref.test(p.0, x, n, alternative="größer")
## B (einseitig größer) = 7.421634 , ( P = 0.00917102 )
```

7.7.11 Die Analyse von Kontingenztafeln

Beispiel Therapieerfolge:

```
erfolg <- matrix(c(14, 22, 32, 18, 16, 8, 8, 2, 0), nr=3, byrow=T,  
  dimnames = list(heilung=c("geheilt-x", "geheilt-x+y", "gestorben"),  
  therapie=c("symptomatisch", "spezifisch N1", "spezifisch N2"))  
as.data.frame(erfolg)
```

	symptomatisch	spezifisch N1	spezifisch N2
geheilt-x	14	22	32
geheilt-x+y	18	16	8
gestorben	8	2	0

```
chisq.test(erfolg, correct = TRUE)  
##  
## Pearson's Chi-squared test  
##  
## data:  erfolg  
## X-squared = 21.576, df = 4, p-value = 0.0002433
```

```
chisq.test(erfolg, simulate.p.value = TRUE, B = 1000)  
##  
## Pearson's Chi-squared test with simulated p-value (based on 1000  
## replicates)  
##  
## data:  erfolg  
## X-squared = 21.576, df = NA, p-value = 0.000999
```

Adjustierte Residuen:

```
n      <- sum(erfolg)
erwartet <- outer(rowSums(erfolg), colSums(erfolg), FUN="*")/n
erwartet <- round(erwartet, 2); erwartet
##               symptomatisch spezifisch N1 spezifisch N2
## geheilt-x      22.67      22.67      22.67
## geheilt-x+y    14.00      14.00      14.00
## gestorben      3.33      3.33      3.33
resid    <- erfolg - erwartet; resid
##               therapie
## heilung        symptomatisch spezifisch N1 spezifisch N2
##   geheilt-x      -8.67      -0.67      9.33
##   geheilt-x+y      4.00      2.00     -6.00
##   gestorben       4.67     -1.33     -3.33
p      <- outer(1-rowSums(erfolg)/n, 1-colSums(erfolg)/n, FUN="*")
adjust  <- resid / sqrt(erwartet * p1); round(adjust, 4)
##               therapie
## heilung        symptomatisch spezifisch N1 spezifisch N2
##   geheilt-x      -2.3910     -0.1848      2.5730
##   geheilt-x+y      1.4037      0.7019     -2.1056
##   gestorben       3.3603     -0.9570     -2.3961
stat    <- sum(resid^2/erwartet); stat
## [1] 21.58584
```

7.7.11.1 Kontingenzkoeffizient (Cramer)

```
V.Cramer <- function(tab) {  
  n      <- sum(tab)  
  nrows <- rowSums(tab); r <- length(nrows)  
  ncols <- colSums(tab); c <- length(ncols);  sum  <- 0  
  for (i in 1:r) {  
    for (j in 1:c) sum <- sum + tab[i,j]^2 / (nrows[i]*ncols[j]) }  
  stat <- n * (sum - 1); stat; pval <- 1- pchisq(stat, df=(r-1)*(c-1))  
  V <- sqrt(stat / (n * min((r-1), c-1)))  
  cat("Chiquadrat =", stat, ", FG =", (r-1)*(c-1), ",  
  P-Wert =", pval, "\n", "Kontingenzkoeffizient nach Cramer V: =", V, "\n")  
}
```

Beispiel Autotypen:

```
cartyp <- matrix(c( 5, 40, 50, 5, 15,  5,  7, 23), nrow=2,  
                 ncol=4, byrow=TRUE,  
                 dimnames=list(job=c("Arbeiter","Angestellter"),  
                                typ=c("Cabrio", "Coupe","Kombi","SUV")))  
as.data.frame(cartyp)
```

	Cabrio	Coupe	Kombi	SUV
Arbeiter	5	40	50	5
Angestellter	15	5	7	23

```
V.Cramer(cartyp)  
## Chiquadrat = 67.01128 , FG = 3 ,  
## P-Wert = 1.865175e-14  
## Kontingenzkoeffizient nach Cramer V: = 0.6683875  
  
chisq.test(cartyp)  
##  
## Pearson's Chi-squared test  
##  
## data:  cartyp  
## X-squared = 67.011, df = 3, p-value = 1.862e-14
```

7.7.11.2 Fallzahl und Power

```
npow.chisq <- function(w=NULL, n=NULL, r=NULL, c=NULL, alpha=NULL, power=NULL) {
  nu <- (r-1)*(c-1)
  quant <- qchisq(alpha, df=nu, lower=FALSE)
  p.expr <- quote(pchisq(quant, df=nu, ncp = n * w^2, lower=FALSE))
  if (is.null(power)) power <- eval(p.expr)
  if (is.null(n))
    n <- uniroot(function(n) eval(p.expr)-power, c(1e-10, 1e+3))$root
  cat("\n", "Fallzahl und Power zum Chiquadrat-Test:", "\n",
      "Effektindex (w)...:", w, "\n",
      "Freiheitsgrade...:", nu, "\n",
      "Signifikanzniveau:", alpha, "\n",
      "Anzahl n (gesamt):", ceiling(n), "\n",
      "Power..... :", power, "\n")
}

npow.chisq(w=0.3, r=2, c=3, alpha=0.05, power=0.80)
##
## Fallzahl und Power zum Chiquadrat-Test:
## Effektindex (w)...: 0.3
## Freiheitsgrade...: 2
## Signifikanzniveau: 0.05
## Anzahl n (gesamt): 108
## Power..... : 0.8

npow.chisq(w=0.3, r=2, c=3, alpha=0.05, n=108)
##
## Fallzahl und Power zum Chiquadrat-Test:
## Effektindex (w)...: 0.3
## Freiheitsgrade...: 2
## Signifikanzniveau: 0.05
## Anzahl n (gesamt): 108
## Power..... : 0.8036939
```

Funktion `pwr.chisq.test()` in `library(pwr)`

```
library(pwr)
pwr.chisq.test(w=0.3, df=(2-1)*(3-1), sig.level=0.05, power=0.80)
##
## Chi squared power calculation
##
## w = 0.3
## N = 107.0521
## df = 2
## sig.level = 0.05
## power = 0.8
##
## NOTE: N is the number of observations
```

7.7.12 Bowker-Test auf Symmetrie

```
bowker.test <- function(tab, k) {  
  stat <- 0  
  for (j in 1:(k-1)) {  
    for (i in (j+1):k) {  
      stat <- stat + ((tab[i,j] - tab[j,i])^2 / (tab[i,j] + tab[j,i]))  
    }  
  }  
  pval <- pchisq(stat, df=k*(k-1)/2, lower.tail = FALSE)  
  cat("Teststatistik:",stat,"(Signifikanz ",pval,")","\n") }  
}
```

Beispiel:

```
k <- 4  
tab <- matrix(c(15,4,6,1, 16,10,3,1, 10,2,4,4, 0,4,12,8),  
             nrow=k, byrow=T); tab  
##      [,1] [,2] [,3] [,4]  
## [1,]  15   4   6   1  
## [2,]  16  10   3   1  
## [3,]  10   2   4   4  
## [4,]   0   4  12   8  
  
bowker.test(tab, k)  
## Teststatistik: 15.2 (Signifikanz 0.01875692 )
```

7.7.13 Marginalhomogenitätstest nach Lehmacher

```
lehmacher.test <- function(tab, k) {  
  stat <- rep(NA, k)  
  r.sum <- apply(tab, 1, sum); c.sum <- apply(tab, 2, sum)  
  for (i in 1:k) stat[i] <- (r.sum[i] - c.sum[i])^2 /  
    (r.sum[i] + c.sum[i] - 2*tab[i,i])  
  p.val <- round(pchisq(stat, df=1, lower.tail = FALSE), 6)  
  p.cor <- round(p.adjust(p.val, "hochberg"), 6)  
  cat("Teststatistik:",round(stat, 4),"\n",  
      "Signifikanz :",round(p.val,4),"\n",  
      "P-adjustiert :",round(p.cor, 4),"\n") }  
}  
  
wahl <- matrix(c(400,40,20,10, 50,300,60,20, 10,40,120,5, 5,90,50,80),  
             nrow=4, byrow=T, dimnames =  
             list(c("Partei A", "Partei B","Partei C","Partei D"),  
                  c("Partei A", "Partei B","Partei C","Partei D")))  
wahl  
##      Partei A Partei B Partei C Partei D  
## Partei A    400     40     20     10  
## Partei B     50    300     60     20  
## Partei C     10     40    120      5  
## Partei D      5     90     50     80  
  
lehmacher.test(wahl, 4)
```

```
## Teststatistik: 0.1852 5.3333 30.4054 67.2222
## Signifikanz : 0.667 0.0209 0 0
## P-adjustiert : 0.667 0.0418 0 0
bowker.test(wahl, 4)
## Teststatistik: 91.47475 (Signifikanz 1.496264e-17 )
```

Beispiel Wahlergebnisse:

```
wahl <- matrix(c(400,40,20,10, 50,300,60,20, 10,40,120,5, 5,90,50,80),
  nrow=4, byrow=T, dimnames =
  list(c("Partei A", "Partei B","Partei C","Partei D"),
       c("Partei A", "Partei B","Partei C","Partei D")))
as.data.frame(wahl)
```

	Partei A	Partei B	Partei C	Partei D
Partei A	400	40	20	10
Partei B	50	300	60	20
Partei C	10	40	120	5
Partei D	5	90	50	80

```
lehmacher.test(wahl, 4)
## Teststatistik: 0.1852 5.3333 30.4054 67.2222
## Signifikanz : 0.667 0.0209 0 0
## P-adjustiert : 0.667 0.0418 0 0

bowker.test(wahl, 4)
## Teststatistik: 91.47475 (Signifikanz 1.496264e-17 )
```

7.7.14 Stuart-Maxwell-Test auf Homogenität in den Randverteilungen

```
stuart.maxwell.test <- function(tab) {
  if(nrow(tab)!=3 | ncol(tab)!=3) {                                # Prüfe Differenzen
    print("Dimension der Tabelle nicht 3*3"); break }
  rs <- rowSums(tab); cs <- colSums(tab)
  d1 <- rs[1] - cs[1]                                              # Randverteilungen
  d2 <- rs[2] - cs[2]
  d3 <- rs[3] - cs[3]
  n12 <- (tab[1,2] + tab[2,1])/2                                    # Symmetriefelder
  n13 <- (tab[1,3] + tab[3,1])/2
  n23 <- (tab[2,3] + tab[3,2])/2
  stat <- (n23*d1^2+n13*d2^2+n12*d3^2)/(2*(n12*n13+n12*n23+n13*n23))
  pval <- pchisq(stat, df=2, lower.tail = FALSE)
  cat("Teststatistik:",stat,"(Signifikanz ",pval,")","\n") }
}
```

Beispiel psychiatrische Störungen:

```
tab <- matrix(c(35,5,0, 15,20,5, 10,5,5),
              nrow=3, byrow=T, dimnames=list(P1=c("D1","D2","D3"),
                                              P2=c("D1","D2","D3")))
as.data.frame(addmargins(tab))
```

	D1	D2	D3	Sum
D1	35	5	0	40
D2	15	20	5	40
D3	10	5	5	20
Sum	60	30	10	100

```
stuart.maxwell.test(tab)
## Teststatistik: 14 (Signifikanz 0.000911882 )
```

Funktion stuart.maxwell.mh() in library(irr)

```
library(irr)
stuart.maxwell.mh(tab)
## Stuart-Maxwell marginal homogeneity
##
## Subjects = 100
## Raters = 2
## Chisq = 14
##
## Chisq(2) = 14
## p-value = 0.000912
```

Funktion `mh_test()` in `library(coin)`

```
library(coin)
mh_test(as.table(tab), distribution=approximate(B=9999))
##
## Approximative Marginal Homogeneity Test
##
## data: response by
## conditions (P1, P2)
## stratified by block
## chi-squared = 14, p-value = 3e-04
```

7.7.15 Q-Test nach Cochran

```
cochranq.test <- function(mat) {
  k <- ncol(mat);          C <- sum(colSums(mat)^2);
  R <- sum(rowSums(mat)^2); T <- sum(rowSums(mat))
  Q <- (k - 1)*((k*C) - (T^2)) / (k*T - R)
  df <- k - 1; names(df) <- "FG"; names(Q) <- "Cochran's Q"
  p.val <- pchisq(Q, df, lower = FALSE)
  QVAL <- list(statistic = Q, parameter = df, p.value = p.val,
               method = "Cochran's Q Test for Dependent Samples",
               data.name = deparse(substitute(mat)))
  class(QVAL) <- "htest"
  return(QVAL)
}
```

Beispiel Expertenurteil zu Weinen:

```
wein <- as.table(matrix(c(1,0,1,1,0, 1,1,1,0,1, 0,0,1,1,1, 1,0,1,0,0,
                        0,0,0,1,1, 1,0,1,1,0), byrow=TRUE, ncol=5,
                        dimnames = list("Person"=c("1","2","3","4","5","6"),
                                         "Wein" = c("A","B","C","D","E"))))

cochranq.test(wein)
##
## Cochran's Q Test for Dependent Samples
##
## data: wein
## Cochran's Q = 5.4118, FG = 4, p-value = 0.2476
```

7.7.16 Cohens's Kappa-Koeffizient

Funktion Kappa() in library(vcd)

```
library(vcd)
attention <- matrix(c(14, 3, 5, 18), nrow=2, ncol=2, byrow=TRUE)
attention
##      [,1] [,2]
## [1,]  14   3
## [2,]   5  18

Kappa(attention)
##           value    ASE      z Pr(>|z|)
## Unweighted 0.597 0.1267 4.711 2.459e-06
## Weighted   0.597 0.1267 4.711 2.459e-06

confint(Kappa(attention))
##
## Kappa          lwr      upr
## Unweighted 0.3486363 0.8453184
## Weighted   0.3486363 0.8453184
```

Hinweis zu Konfidenzintervallen (Bootstrap):

Funktion ckappa() in library(psy)

```
library(psy); library(boot)
b1 <- c(rep(0,17), rep(1,23)); b2 <- c(rep(0,14),
                                     rep(1,3), rep(0,5), rep(1,18))
attention <- as.data.frame(cbind(b1, b2))

ckappa(attention)$kappa
## [1] 0.5969773

ckappa.boot <- function(data, x) {ckappa(data[x,])[[2]]}
res <- boot(attention, ckappa.boot, 500)

boot.ci(res, type="bca")
## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
## Based on 500 bootstrap replicates
##
## CALL :
## boot.ci(boot.out = res, type = "bca")
##
## Intervals :
## Level      BCa
## 95%      ( 0.3052, 0.8000 )
## Calculations and Intervals on Original Scale
## Some BCa intervals may be unstable
```

7.7.16.1 Das gewichtete Kappa

Beispiel Botulinum:

Funktion Kappa() in library(vcd)

```
botulin <- matrix(c(5,2,0,1,0, 1,7,2,2,0, 1,2,10,5,1,
                    0,0,3,4,0, 0,0,0,0,3),
                  nrow=5, ncol=5, byrow=TRUE,
                  dimnames = list("U1"=c("0","I","II","III","IV"),
                                   "U2"=c("0","I","II","III","IV")))
as.data.frame(botulin)
```

	0	I	II	III	IV
0	5	2	0	1	0
I	1	7	2	2	0
II	1	2	10	5	1
III	0	0	3	4	0
IV	0	0	0	0	3

```
library(vcd)
Kappa(botulin, weights = "Fleiss-Cohen")
##           value      ASE      z  Pr(>|z|)
## Unweighted 0.4651 0.09339 4.980 6.370e-07
## Weighted   0.6849 0.09569 7.158 8.216e-13

confint(Kappa(botulin, weights = "Fleiss-Cohen"))
##
## Kappa          lwr          upr
## Unweighted 0.2820184 0.6481126
## Weighted   0.4973423 0.8724326
```

7.7.16.2 Das Kappa für mehrfache Beurteilungen (Multi-Rater)

Bewertung von Röntgenaufnahmen:

```
radiol <- matrix(c(1,4,0, 2,0,3, 0,0,5, 4,0,1, 3,0,2,
                  1,4,0, 5,0,0, 0,4,1, 1,0,4, 3,0,2),
                  nrow=10, ncol=3, byrow=TRUE)

n      <- 10; R <- 5; k <- 3;
p.i    <- rep(NA, n);

for (i in 1:n) p.i[i] <- sum(radiol[i,]*(radiol[i,]-1))/(R*(R-1))
p.bar <- sum(p.i)/n; p.bar
## [1] 0.62

p.j    <- rep(NA, k); for (j in 1:k) p.j[j] <- sum(radiol[,j])/(n*R)
p.e    <- sum(p.j^2); p.e
```

```
## [1] 0.3472

kappa.m <- (p.bar - p.e)/(1-p.e)
kappa.m
## [1] 0.4178922

var <- (2/(n*R*(R-1))) * (p.e-(2*R-3)*p.e^2+2*(R-2)*sum(p.j^3))/(1-p.e)^2
var
## [1] 0.005872261

z <- kappa.m / sqrt(var)
z
## [1] 5.453327

2*pnorm(z, lower.tail=FALSE)
## [1] 4.943598e-08
```

Funktion kappam.fleiss() in library(irr)

```
library(irr)
data <- matrix(c(1,2,2,2,2, 1,1,3,3,3, 3,3,3,3,3, 1,1,1,1,3, 1,1,1,3,3,
                 1,2,2,2,2, 1,1,1,1,1, 2,2,2,2,3, 1,3,3,3,3, 1,1,1,3,3),
               nrow=10, byrow=T,
               dimnames=list(Bild=1:10,
                             Untersucher=c("U1", "U2", "U3", "U4", "U5")))

kappam.fleiss(data, exact = FALSE, detail = FALSE)
## Fleiss' Kappa for m Raters
##
## Subjects = 10
## Raters = 5
## Kappa = 0.418
##
## z = 5.83
## p-value = 5.47e-09
```

7.7.17 Krippendorff's Alpha

Beispiel Reliabilität mit Funktion `kripp.alpha()` in `library(irr)`:

```
o1 <- c(1, 2, 3, 3, 2, 1, 4, 1, 2, NA, NA, NA)
o2 <- c(1, 2, 3, 3, 2, 2, 4, 1, 2, 5, NA, 3)
o3 <- c(NA, 3, 3, 3, 2, 3, 4, 2, 2, 5, 1, NA)
o4 <- c(1, 2, 3, 3, 2, 4, 4, 1, 2, 5, 1, NA)
dat <- rbind(Obs_A=o1, Obs_B=o2, Obs_C=o3, Obs_D=o4)
dat
##           [,1] [,2] [,3] [,4] [,5] [,6] [,7] [,8] [,9] [,10] [,11] [,12]
## Obs_A      1   2   3   3   2   1   4   1   2   NA   NA   NA
## Obs_B      1   2   3   3   2   2   4   1   2   5   NA   3
## Obs_C     NA   3   3   3   2   3   4   2   2   5   1   NA
## Obs_D      1   2   3   3   2   4   4   1   2   5   1   NA

library(irr)
kripp.alpha(dat, method="ordinal")
## Krippendorff's alpha
##
## Subjects = 12
## Raters = 4
## alpha = 0.815
```

7.7.18 Kendall's Konkordanzkoeffizient

Beispiel Schönheitswettbewerb mit Funktion `kendall()` in `library(irr)`

```
library(irr)
ranking <- matrix(c(3,1,4,6,5,7,8,2,
                    2,3,5,7,4,6,8,1,
                    3,2,5,4,6,8,7,1), nrow=3, byrow=T)
kendall(t(ranking), correct=TRUE)
## Kendall's coefficient of concordance Wt
##
## Subjects = 8
## Raters = 3
## Wt = 0.894
##
## Chisq(7) = 18.8
## p-value = 0.00891

pchisq(18.8, 7, ncp=0, lower.tail = F, log.p = FALSE)
## [1] 0.008837491
```

7.8 Hypothesentests zur Korrelation und Regression

7.8.1 Korrelationskoeffizient nach Pearson

```
x <- c( 4 , 6 , 8 , 3 , 9 , 5 , 10 , 2 , 7 , 8)
y <- c( 5 , 5 , 7 , 4 , 11 , 7 , 9 , 5 , 8 , 10)
n <- length(x)

r <- cor(x, y, method="pearson"); r
## [1] 0.8401183

t_hat <- r * sqrt((n-2)/(1-r^2)); t_hat
## [1] 4.380899

cor.test(x, y, method="pearson")
##
## Pearson's product-moment correlation
##
## data:  x and y
## t = 4.3809, df = 8, p-value = 0.002346
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
##  0.4468671 0.9612704
## sample estimates:
##          cor
## 0.8401183
```

Fisher-Transformation von r auf z:

```
r <- seq(0, 0.99, 0.01); zp <- function(r) 0.5*log((1+r)/(1-r))
tab <- matrix(round(zp(r),4), byrow=T, nrow=10);
colnames(tab) <- seq(0.00, 0.09, 0.01)
rownames(tab) <- seq(0.0, 0.9, 0.1)
as.data.frame(tab[1:10,])
```

	0	0.01	0.02	0.03	0.04	0.05	0.06	0.07	0.08	0.09
0	0.0000	0.0100	0.0200	0.0300	0.0400	0.0500	0.0601	0.0701	0.0802	0.0902
0.1	0.1003	0.1104	0.1206	0.1307	0.1409	0.1511	0.1614	0.1717	0.1820	0.1923
0.2	0.2027	0.2132	0.2237	0.2342	0.2448	0.2554	0.2661	0.2769	0.2877	0.2986
0.3	0.3095	0.3205	0.3316	0.3428	0.3541	0.3654	0.3769	0.3884	0.4001	0.4118
0.4	0.4236	0.4356	0.4477	0.4599	0.4722	0.4847	0.4973	0.5101	0.5230	0.5361
0.5	0.5493	0.5627	0.5763	0.5901	0.6042	0.6184	0.6328	0.6475	0.6625	0.6777
0.6	0.6931	0.7089	0.7250	0.7414	0.7582	0.7753	0.7928	0.8107	0.8291	0.8480
0.7	0.8673	0.8872	0.9076	0.9287	0.9505	0.9730	0.9962	1.0203	1.0454	1.0714
0.8	1.0986	1.1270	1.1568	1.1881	1.2212	1.2562	1.2933	1.3331	1.3758	1.4219
0.9	1.4722	1.5275	1.5890	1.6584	1.7380	1.8318	1.9459	2.0923	2.2976	2.6467

Fisher-Transformtion von z auf r:

```
z <- seq(0, 3, 0.01); zr <- function(z) (exp(2*z)-1)/(exp(2*z)+1)
tab <- matrix(round(zr(z), 4), byrow=T, ncol=10)
colnames(tab) <- seq(0.00, 0.09, 0.01)
rownames(tab) <- seq(0.0, 3, 0.1)
as.data.frame(tab[1:10,])
```

	0	0.01	0.02	0.03	0.04	0.05	0.06	0.07	0.08	0.09
0	0.0000	0.0100	0.0200	0.0300	0.0400	0.0500	0.0599	0.0699	0.0798	0.0898
0.1	0.0997	0.1096	0.1194	0.1293	0.1391	0.1489	0.1586	0.1684	0.1781	0.1877
0.2	0.1974	0.2070	0.2165	0.2260	0.2355	0.2449	0.2543	0.2636	0.2729	0.2821
0.3	0.2913	0.3004	0.3095	0.3185	0.3275	0.3364	0.3452	0.3540	0.3627	0.3714
0.4	0.3799	0.3885	0.3969	0.4053	0.4136	0.4219	0.4301	0.4382	0.4462	0.4542
0.5	0.4621	0.4699	0.4777	0.4854	0.4930	0.5005	0.5080	0.5154	0.5227	0.5299
0.6	0.5370	0.5441	0.5511	0.5581	0.5649	0.5717	0.5784	0.5850	0.5915	0.5980
0.7	0.6044	0.6107	0.6169	0.6231	0.6291	0.6351	0.6411	0.6469	0.6527	0.6584
0.8	0.6640	0.6696	0.6751	0.6805	0.6858	0.6911	0.6963	0.7014	0.7064	0.7114
0.9	0.7163	0.7211	0.7259	0.7306	0.7352	0.7398	0.7443	0.7487	0.7531	0.7574

7.8.1.2 Korrelation bei Mehrfachbeobachtungen

Beispieldatensatz nach J.M. Bland:

```
daten <- read.csv2("blandcor.csv")
subj <- 1:4

attach(daten)
cor.test(y, x) # Pearson Korrelation
##
## Pearson's product-moment correlation
##
## data: y and x
## t = 4.3809, df = 8, p-value = 0.002346
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## 0.4468671 0.9612704
## sample estimates:
## cor
## 0.8401183

tmp <- by(daten, id, function(d) cor(d$y, d$x))
korr <- tmp[1:4]; round(korr,3) # Einzelfälle
## id
## 1 2 3 4
## -0.333 0.486 0.065 -0.389

tmp.y <- by(daten, id, function(d) mean(d$y))
tmp.x <- by(daten, id, function(d) mean(d$x))
tmp.n <- by(daten, id, function(d) length(d$x))
cor.test(tmp.y, tmp.x) # Korrelation aus Mittelwerten
```

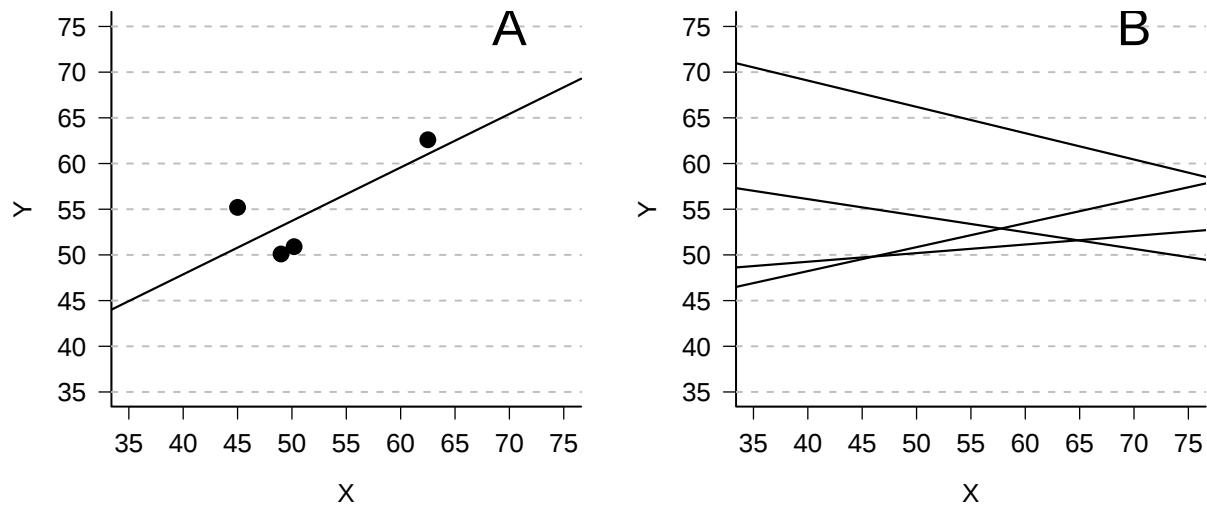
```
##
## Pearson's product-moment correlation
##
## data: tmp.y and tmp.x
## t = 1.7177, df = 2, p-value = 0.228
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## -0.7327773 0.9949069
## sample estimates:
## cor
## 0.7720022

mx <- round(tmp.x, 2); my <- round(tmp.y, 2); w <- tmp.n
korr.w <- (sum(w*mx*my) - sum(w*mx)*sum(w*my) / sum(w)) /
  sqrt( (sum(w*mx^2) - (sum(w*mx)^2/sum(w))) *
        (sum(w*my^2) - (sum(w*my)^2/sum(w))) )
korr.w # Pearson Korrelation gewichtet
## [1] 0.7720022

par(mfcol=c(1,2), lwd=1.5, font.axis=1.5, bty="l", ps=15)
plot(x[id==subj[1]], y[id==subj[1]], las=1,
     xlab="X", ylab="Y", xlim=c(35, 75), ylim=c(35, 75),
     xaxp=c(35, 75, 8), yaxp=c(35, 75, 8), cex=1.0, pch=1)
for (i in 2:4) points(x[id==subj[i]], y[id==subj[i]], cex=1.0, pch=i)
abline(h=seq(35, 75, 5), lty=2, col="grey")
text(70,75,"A", cex=2)
points(tmp.x, tmp.y, pch=16, cex=1.8) # Regression zu Mittelwerten
abline(lm(tmp.y ~ tmp.x), lty=1, lwd=1.7)

plot(x[id==subj[1]], y[id==subj[1]], las=1,
     xlab="X", ylab="Y", xlim=c(35, 75), ylim=c(35, 75),
     xaxp=c(35, 75, 8), yaxp=c(35, 75, 8), cex=1.5, pch=1)
for (i in 2:4) points(x[id==subj[i]], y[id==subj[i]], cex=1.5, pch=i)
abline(h=seq(35, 75, 5), lty=2, col="grey")
# Regression zu einzelnen Faellen

tmp <- with(daten, by(daten, id, function(d) lm(y ~ x, data = d)))
regr <- sapply(tmp, coef)
for (i in 1:4) abline(a=regr[1,i], b=regr[2,i], lty=1, lwd=1.7)
text(70,75,"B", cex=2)
```



Korrelation innerhalb der Fälle:

```
vartab <- summary(aov(y ~ as.factor(id) + x, data=daten))
vartab
##              Df Sum Sq Mean Sq F value    Pr(>F)
## as.factor(id)  3  982.6   327.5   26.317 4.34e-09 ***
## x              1    0.2     0.2    0.016   0.901
## Residuals     35  435.6    12.4
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

saq <- as.vector(vartab[[1]][2])
korr.w <- sqrt(saq[2,1] / (saq[2,1]+saq[3,1]))
korr.w
## [1] 0.02118205
```

7.8.1.3 Fallzahl und Power zum Korrelationskoeffizienten

```
npwr.rho <- function(n=NULL, r, sig.lev=0.05, power=NULL) {
  if (sum(sapply(list(n, power), is.null)) != 1)
    stop("nur n oder power dürfen fehlen")
  z.alpha <- qnorm(1-sig.lev/2) # nur zweiseitige Hypothesenstellung
  if (is.null(n)) {
    z.beta <- qnorm(power)
    n <- ((z.alpha + z.beta)/atanh(r))^2 + 3
    return(ceiling(n))
  }
  if (is.null(power)) {
    z.beta <- sqrt(n-3)*atanh(r) - z.alpha
    power <- pnorm(z.beta)
    return(round(power, 4))
  }
}
```

```
npwr.rho(r=0.60, power=0.90)
## [1] 25
```

Tabelle:

```
rho <- seq(0.1, 0.9, by=0.1); pwr <- seq(0.5, 0.9, by=0.1)
tab <- matrix(rep(0, 5*9), byrow=T, nrow=9)

for (i in 1:5) tab[,i] <- npwr.rho(r=rho, power=pwr[i], sig.lev=0.05) # Alpha = 0.05
colnames(tab) <- seq(0.50, 0.90, 0.10)
rownames(tab) <- seq(0.10, 0.90, 0.10)
as.data.frame(tab)
```

	0.5	0.6	0.7	0.8	0.9
0.1	385	490	617	783	1047
0.2	97	123	154	194	259
0.3	44	55	68	85	113
0.4	25	31	38	47	62
0.5	16	20	24	30	38
0.6	11	14	16	20	25
0.7	9	10	12	14	17
0.8	7	8	9	10	12
0.9	5	6	6	7	8

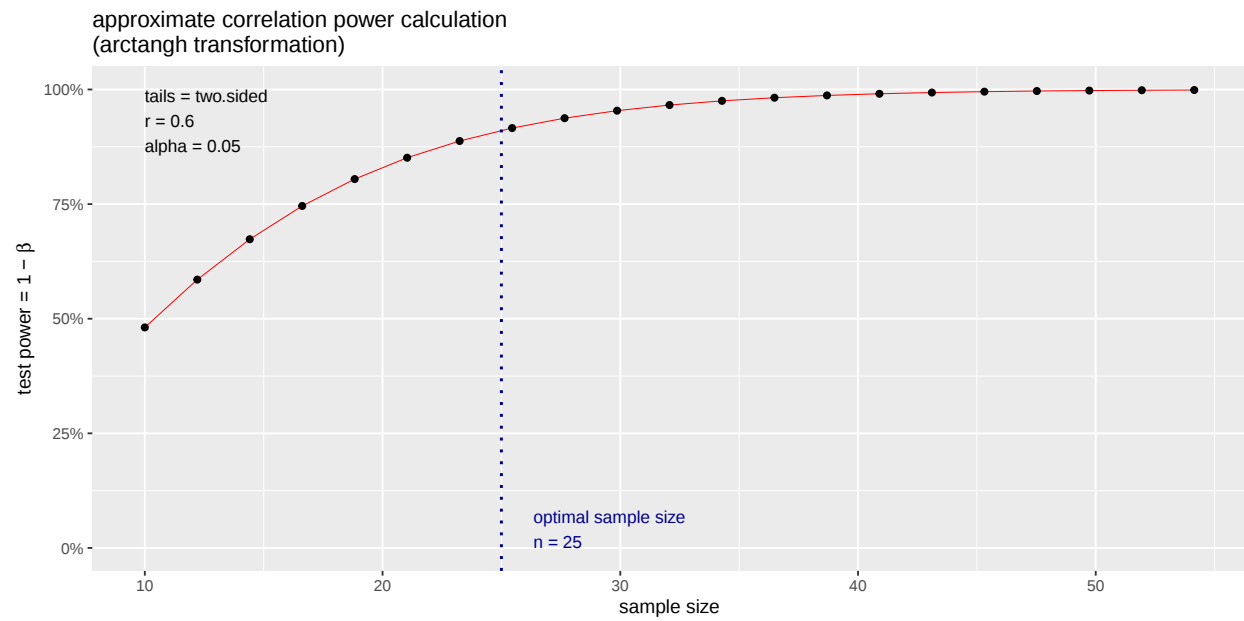
```
for (i in 1:5) tab[,i] <- npwr.rho(r=rho, power=pwr[i], sig.lev=0.01) # Alpha = 0.01
colnames(tab) <- seq(0.50, 0.90, 0.10)
rownames(tab) <- seq(0.10, 0.90, 0.10)
as.data.frame(tab)
```

	0.5	0.6	0.7	0.8	0.9
0.1	663	799	958	1164	1482
0.2	165	198	237	288	366
0.3	73	87	104	125	159
0.4	40	48	57	69	86
0.5	25	30	35	42	53
0.6	17	20	24	28	34
0.7	12	14	16	19	23
0.8	9	10	11	13	16
0.9	7	7	8	9	10

Funktion `pwr.r.test()` in `library(pwr)`

```
library(pwr)
prp <- pwr.r.test(n=NULL, r=0.60, power=0.90, alternative="two.sided")
prp
##
##      approximate correlation power calculation (arctangh transformation)
##
```

```
##          n = 24.14439
##          r = 0.6
##      sig.level = 0.05
##          power = 0.9
##  alternative = two.sided
plot(prp)
```



7.8.2 Prüfung der Rangkorrelationskoeffizienten

Schranken für Spearman und Kendall:

Funktion `q.Kendall()` und `q.Spearman()` in `library(SuppDists)`

```
library(SuppDists) # einseitige obere Schranken

alpha <- c(0.005, 0.025, 0.05); q <- 1 - alpha
n     <- 4:30

q.Kendall <- matrix(NA, nrow=27, ncol=3)
q.Spearman <- matrix(NA, nrow=27, ncol=3)

for (i in 4:30) {
  for(j in 1:3) {
    q.Kendall[i-3, j] <- qKendall(q[j], n[i-3], lower.tail=TRUE)
    q.Spearman[i-3, j] <- qSpearman(q[j], n[i-3], lower.tail=TRUE)
  } }
tab <- cbind(round(q.Spearman, 3), round(q.Kendall, 3))
colnames(tab) <- c("Spearman 0.01", "Spearman 0.05", "Spearman 0.10",
                  "Kendall 0.01", "Kendall 0.05", "Kendall 0.10")
rownames(tab) <- 4:30
as.data.frame(tab)
```

	Spearman 0.01	Spearman 0.05	Spearman 0.10	Kendall 0.01	Kendall 0.05	Kendall 0.10
4	1.000	1.000	0.800	1.000	1.000	0.667
5	1.000	0.900	0.800	1.000	0.800	0.600
6	1.000	0.943	0.829	0.867	0.733	0.600
7	0.929	0.821	0.750	0.810	0.619	0.524
8	0.881	0.762	0.667	0.714	0.571	0.500
9	0.833	0.700	0.617	0.667	0.500	0.444
10	0.782	0.648	0.576	0.600	0.467	0.422
11	0.755	0.618	0.536	0.564	0.455	0.382
12	0.720	0.587	0.507	0.545	0.424	0.364
13	0.692	0.560	0.484	0.538	0.410	0.359
14	0.670	0.538	0.464	0.495	0.385	0.341
15	0.645	0.521	0.446	0.486	0.371	0.314
16	0.626	0.503	0.432	0.467	0.367	0.300
17	0.610	0.485	0.417	0.456	0.353	0.294
18	0.593	0.472	0.404	0.438	0.333	0.281
19	0.578	0.458	0.391	0.427	0.322	0.275
20	0.564	0.447	0.380	0.411	0.316	0.263
21	0.550	0.434	0.371	0.400	0.305	0.257
22	0.539	0.425	0.362	0.385	0.299	0.255
23	0.528	0.415	0.353	0.383	0.289	0.249
24	0.516	0.406	0.345	0.370	0.283	0.239
25	0.506	0.398	0.338	0.360	0.280	0.233
26	0.497	0.389	0.331	0.354	0.274	0.231
27	0.487	0.382	0.324	0.350	0.265	0.225
28	0.479	0.375	0.318	0.339	0.259	0.222
29	0.471	0.368	0.312	0.335	0.256	0.217
30	0.464	0.362	0.307	0.329	0.251	0.214

Beispiel Weinsorten - Spearman:

```
x <- c(1, 2, 3, 4, 5, 6, 7, 8, 9, 10)
y <- c(2, 1, 5, 3, 4, 6, 7, 9, 8, 10)

cor.test(x, y, method="spearman")
##
## Spearman's rank correlation rho
##
## data: x and y
## S = 10, p-value < 2.2e-16
## alternative hypothesis: true rho is not equal to 0
## sample estimates:
## rho
## 0.9393939
```

Beispiel Weinsorten - Kendall:

```
x <- c(1, 2, 3, 4, 5, 6, 7, 8, 9, 10)
y <- c(2, 1, 5, 3, 4, 6, 7, 9, 8, 10)

cor.test(x, y, method="kendall")
##
## Kendall's rank correlation tau
##
## data: x and y
## T = 41, p-value = 0.0003577
## alternative hypothesis: true tau is not equal to 0
## sample estimates:
## tau
## 0.8222222
```

7.8.4 Hypothesentests zu den Regressionskoeffizienten

7.8.4.1 Prüfung der Linearität

```
xi <- c(1, 5, 9, 13); k <- length(xi)
ni <- c(2, 3, 1, 2); n <- sum(ni)
yij <- matrix(c(1,2,NA, 2,3,3, 4,NA,NA, 5,6,NA), ncol=k, byrow=FALSE)

yisum <- rep(0, k) # Gruppenmittelwerte
for (j in 1:k) {for (i in 1:ni[j]) yisum[j] <- yisum[j] + yij[i,j]}
yibar <- yisum / ni # lineare Regression (x, y)

x <- NULL; for (j in 1:k) x <- c(x, rep(xi[j], ni[j]))
y <- NULL; for (j in 1:k) {for (i in 1:ni[j]) y <- c(y, yij[i,j])}
linreg <- lm(y~x); a <- linreg$coeff[1]; b <- linreg$coeff[2]
yihat <- a + b*xi # Schätzung

ZF <- (1/(k-2))*sum(ni*(yibar-yihat)^2) # Abweichungen
sn <- 0 # vom Gruppenmittelwert
for (j in 1:k) {for (i in 1:ni[j]) sn <- sn + (yij[i,j] - yibar[j])^2}
NF <- (1/(n-k))*sn

F <- ZF/NF; F # Teststatistik F
## [1] 0.06582278
```

7.8.4.2 Chow-Test: Strukturbruch

```
chow.test <- function(x, y, gruppe) {
  n <- length(x); k <- 2
  if (length(y) != n | length(gruppe) != n)
    stop("unterschiedliche Laengen in 'x', 'y' oder 'gruppe' ")
  x1 <- x[gruppe==1]; x2 <- x[gruppe==2]
  y1 <- y[gruppe==1]; y2 <- y[gruppe==2]
  ur.reg <- lm(y ~ x) # lineare Regressionsmodelle lm()
  r.reg1 <- lm(y1 ~ x1); r.reg2 <- lm(y2 ~ x2)
  SSR = NULL # Summe der quadrierten. Residuen
  SSR$ur <- ur.reg$residuals^2; sum(SSR$ur)
  SSR$r1 <- r.reg1$residuals^2; sum(SSR$r1)
  SSR$r2 <- r.reg2$residuals^2; sum(SSR$r2)
  zaehler <- (sum(SSR$ur) - (sum(SSR$r1) + sum(SSR$r2))) / k
  nenner <- (sum(SSR$r1) + sum(SSR$r2)) / (n - 2*k)
  chow <- zaehler / nenner; P <- 1 - pf(chow, k, (n - 2*k))
  cat(sep=" ", "\n", "Chow-Test: F=", chow, " mit ", k, "; ", n-2*k,
      " Freiheitsgraden (P-Wert=", P, ")", "\n")
}
```

Beispiel:

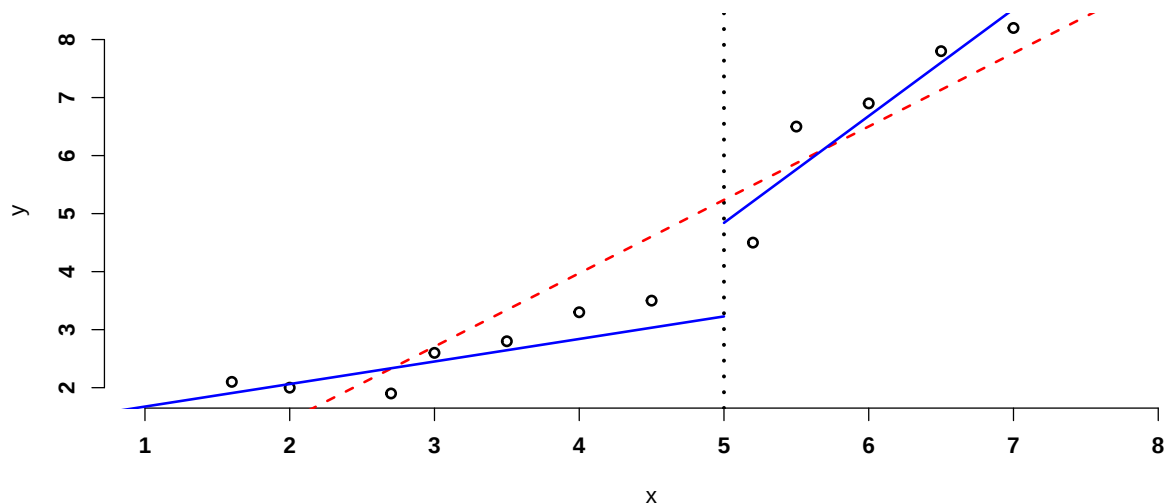
```
x <- c(1.6, 2.0, 2.7, 3.0, 3.5, 4.0, 4.5, 5.2, 5.5, 6.0, 6.5, 7.0)
y <- c(2.1, 2.0, 1.9, 2.6, 2.8, 3.3, 3.5, 4.5, 6.5, 6.9, 7.8, 8.2)

grp <- c(rep(1, 7), rep(2, 5))
chow.test(x, y, grp)
##
## Chow-Test: F=10.72839 mit 2;8 Freiheitsgraden (P-Wert=0.005440255)
```

```
n <- length(x); cut <- 5; ol <- 8
groups <- c(rep(1, cut), rep(2, n-cut))
dfr <- as.data.frame(cbind(x,y,groups))

ur.reg <- lm(y ~ x, data = dfr) # Regression (alle Daten)
r.reg1 <- lm(y ~ x, data = dfr[1:cut,]) # beschränkt auf 1:19
r.reg2 <- lm(y ~ x, data = dfr[cut+1:n,]) # beschränkt auf 20:38

par(mfcol=c(1,1),lwd=2, font.axis=2, bty="n", ps=13)
plot(x, y, xlim=c(1,ol), ylim=range(y))
abline(v=cut, lwd=3, lty=3)
abline(ur.reg, col = "red", lwd = 2, lty = "dashed") # volles Modell
segments(0, r.reg1$coefficients[1], cut, # Modell 1 / 2
  r.reg1$coefficients[1] + cut * r.reg1$coefficients[2], col= 'blue')
segments(cut, r.reg2$coefficients[1] + cut * r.reg2$coefficients[2],
  ol, r.reg2$coefficients[1] + ol * r.reg2$coefficients[2], col= 'blue')
```

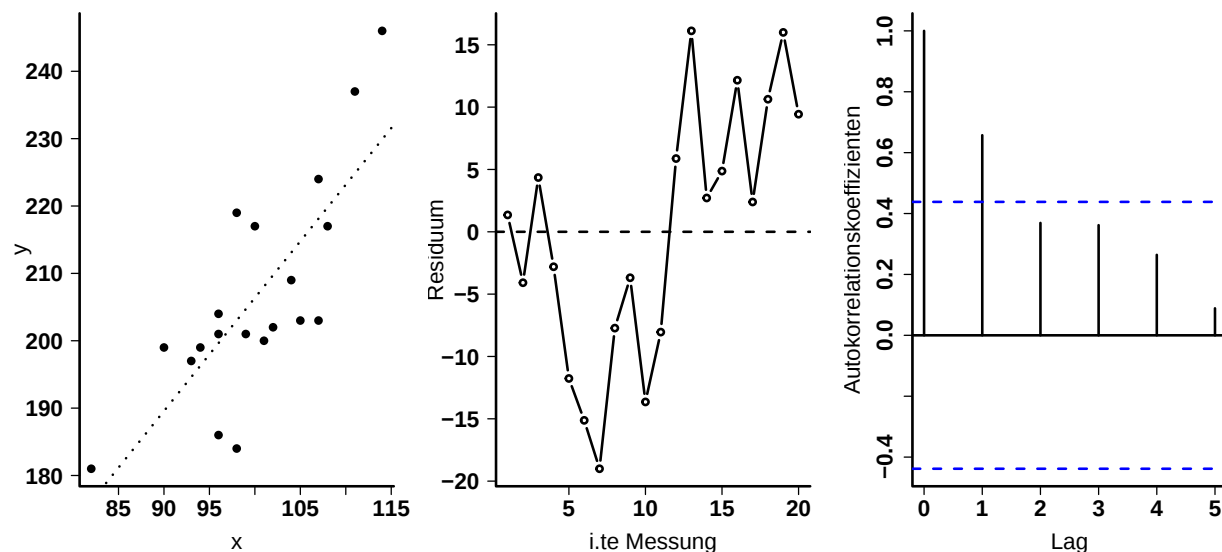


7.8.4.3 Durbin-Watson-Test: Autokorrelation in den Residuen

```
x <- c( 96,104, 96,108,105,107, 98,102, 99, 96,101,
       107,114, 94, 82,111, 93,100, 98, 90)
y <- c(201,209,204,217,203,203,184,202,201,186,200,
       224,246,199,181,237,197,217,219,199)
n <- length(x)

mod <- lm(y~x); e <- residuals(mod); # summary(mod)

par(mfcol=c(1,3), lwd=2, font.axis=2, bty="l", ps=20)
plot(x, y, las=1, xlab="x", ylab="y", cex=1.4, pch=16, col="black")
abline(mod, lty=3, col="black")
plot(1:n, e, type="b", las=1, xlab="i.te Messung", ylab="Residuum")
abline(h=0, lty=2, col="black")
acorr <- acf(e, lag.max=5, main=" ", ylab="Autokorrelationskoeffizienten")
```



```
mod <- lm(y~x); e <- residuals(mod)
d <- rep(NA, n-1)
for (i in 2:n) d[i-1] <- (e[i] - e[i-1])
dw_stat <- sum(d^2)/sum(e^2); dw_stat # Durbin-Watson Test
## [1] 0.6408784
```

Funktion `dwtest()` in library(`lmtest`)

```
library(lmtest)
dwtest(mod)
##
## Durbin-Watson test
##
```

```
## data: mod
## DW = 0.64088, p-value = 0.0001659
## alternative hypothesis: true autocorrelation is greater than 0
```

Korrektur für Autokorrelation:

```
lag_e <- rep(NA, n-1); lag_y <- rep(NA,n-1); lag_x <- rep(NA, n-1)
for (i in 2:n) {lag_e[i] <- e[i-1];
                lag_y[i] <- y[i-1];
                lag_x[i] <- x[i-1] }
mc <- lm(e ~ lag_e - 1)
rho <- mc$coefficients; rho                                     # Autokorrelationskoeffizient
##      lag_e
## 0.6872998
y_c <- y - rho*lag_y;   x_c <- x - rho*lag_x
modc<- lm(y_c ~ x_c); summary(modc)
##
## Call:
## lm(formula = y_c ~ x_c)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -10.3635  -7.1165   0.8942   7.1390   9.0663
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    2.2821     5.8068   0.393   0.699
## x_c            2.0007     0.1783  11.219 2.8e-09 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 7.414 on 17 degrees of freedom
## (1 observation deleted due to missingness)
## Multiple R-squared:  0.881, Adjusted R-squared:  0.874
## F-statistic: 125.9 on 1 and 17 DF, p-value: 2.796e-09
```